



10/23/97

ASST. COMMISSIONER FOR
PATENTS
Washington, D.C. 20231

Docket No.: P-CE 2817

A
Box Sep.

Sir:

Transmitted herewith for filing is the patent application of
Inventor(s): Julie R. Korenberg
For: NUCLEIC ACID ENCODING DS-CAM PROTEINS AND PRODUCTS RELATED THERETO

Enclosed are:

- ☒ 4 sheets of drawing(s), 1 cover page, 1 abstract page,
100 specification pages
☐ An assignment of the invention to _____
☐ A certified copy of a _____ application.
☐ A combination declaration and power of attorney.
☒ Sequence Listing.
☒ Sequence Disk.
☒ Request for Filing Application Under 37 CFR 1.53(a), (b) & (d).
☒ Statement Under 37 CFR §1.821(f).

The filing fee has been calculated as shown below:

	Number Filed		Number Extra		Rate			Fee	
					Small Entity	Other Entity		Small Entity	Other Entity
Total Claims	30 - 20	=	10	x	\$11	\$22	=	\$	\$
Independent Claims	5 - 3	=	2	x	\$41	\$82	=	\$	\$
Multiple Dependent Claims Presented: <input type="checkbox"/> Yes <input checked="" type="checkbox"/> No					\$135	\$270		\$	\$
					BASIC FEE			\$395	\$790
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- ☐ Any patent application processing fees under 37 C.F.R. 1.17.
- ☐ The issue fee set in 37 C.F.R. 1.18 at or before mailing of the Notice of Allowance, pursuant to 37 C.F.R. 1.311(b).
- ☐ Any filing fees under 37 C.F.R. 1.16 for presentation of extra claims.

Respectfully submitted,

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CERTIFICATE OF MAILING BY "EXPRESS MAIL"

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

REQUEST FOR FILING APPLICATION UNDER 37 CFR 1.53(a), (b) & (d)

The Asst. Commissioner for Patents
Washington, D.C. 20231

Ref. No. P-CE 2817

Sir:

This is a Request for filing a new patent application entitled:

NUCLEIC ACID ENCODING DS-CAM PROTEINS AND
PRODUCTS RELATED THERETO

without an Assignment or Oath/Declaration but for which is
enclosed the following:

- [X] Cover Page;
[X] Abstract 1 page(s);
[X] 100 pages of Specification (only spec. and claims);
[] Specification in non-English language;
[X] 30 numbered claim(s); and
[X] 4 sheet(s) of drawings [X] informal; [] formal;
size: [X] A4 [] 14" [] 11"
[] Verified Statement claiming Small Entity Status
(37 CFR 1.9(f) and 1.27(d)) filed December 17, 1996.
[] This application is based on prior foreign application(s)
No.(s). _____ filed in _____ on
_____, respectively, and priority is hereby claimed
therefrom.
[] This application is a [] Continuation [] Divisional
[] Continuation-in-part of application serial no.
_____ filed _____.
[X] This application is based on, and claims the benefit of,
U.S. Provisional Application No. 60/029,322, filed October
25, 1996, entitled NUCLEIC ACID ENCODING A NOVEL DS-CAM
PROTEIN AND PRODUCTS RELATED THERETO.

This application is made by the following named inventor(s):

1. Inventor: Julie R. Korenberg
Citizenship: United States of America
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Post Office Address: 8125 Skyline Drive
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Respectfully submitted,

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Stephen Nguyen
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PATENT

Our Docket: P-CE 2817

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:)
Julie R. Korenberg)
Serial No.: Unknown)
Filed: October 23, 1997)
For: NUCLEIC ACID ENCODING)
DS-CAM PROTEINS AND)
PRODUCTS RELATED THERETO)

Group Art Unit: Unknown

Examiner: Unknown

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Washington, D.C. 20231

Stephane Nguyen
(TYPED OR PRINTED NAME OR PERSON MAILING PAPER OR FEE)

[Signature]
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Sir:

STATEMENT UNDER 37 C.F.R. § 1.821(f)

I hereby state that the content of the paper and
computer readable copies of the Sequence Listing, submitted in
accordance with 37 CFR § 1.821(c) and (e), respectively, are the
same.

Respectfully submitted,

Date : October 23, 1997

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NUCLEIC ACID ENCODING DS-CAM PROTEINS
AND PRODUCTS RELATED THERETO

This is a non-provisional application based on,
and claims the benefit of, U.S. Provisional Application
5 No. 60/029,322 filed October 25, 1996, the content of
which is incorporated herein by reference in its
entirety.

ACKNOWLEDGMENT

This invention was made with Government support
10 under Grant Numbers HL50025 and HD17449 awarded by the
National Institutes of Health and DE-FG03-92ER61402
awarded by the Department of Energy. The Government has
certain rights in this invention.

FIELD OF THE INVENTION

15 The present invention relates to nucleic acids
and proteins encoded thereby. Invention nucleic acids
encode a novel N-CAM member of the immunoglobulin
superfamily of proteins. The invention also relates to
methods for making and using such nucleic acids and
20 proteins.

BACKGROUND OF THE INVENTION

Research spanning the last decade has
significantly elucidated the molecular events attending
cell-cell interactions in the body, especially those
25 events involved in the movement and activation of cells
in the immune system. See generally, Springer et al.,
Nature 346:425-434, 1990. Cell surface proteins, and
especially the so-called Cellular Adhesion Molecules
("CAMs") have correspondingly been the subject of
30 pharmaceutical research and development having as its
goal intervening in the processes of leukocyte

extravasation to sites of inflammation and leukocyte movement to distinct target tissues. The isolation and characterization of cellular adhesion molecules, the cloning and expression of DNA sequences encoding such molecules, and the development of therapeutic and diagnostic agents relevant to inflammatory process, viral infection and cancer metastasis have also been the subject of numerous U.S. and foreign applications for Letters Patent. See Edwards, Current Opinion in Therapeutic Patents 1(11):1617-1630, 1991 and particularly the published "patent literature references" cited therein.

Numerous CAMs have been characterized to date. See, for example, vascular adhesion molecule (VCAM-1) as described in PCT WO 90/13300; platelet endothelial cell adhesion molecule (PECAM-1) described in Newman et al., Science 247:1219-1222, 1990; and PCT WO 91/10683; and the following U.S. Patents: 5,525,487; 5,235,049; 5,272,263; 5,489,233; 5,264,554; 5,318,890; 5,389,520; 5,519,008; and the like.

There is substantial evidence that N-CAM and its relatives play an important part in neural development (Edelman and Crossin, "CELL ADHESION MOLECULES: Implications for a Molecular Histology", Ann. Rev. Biochem. 60:155-190, 1991; and Walsh and Doherty, Curr. Opinion in Cell Biol. 5:791-796, 1993). For example, antibodies directed against N-CAMs disturbed the normal growth pattern of nerve processes. N-CAM (locus 11q23.1) is expressed in large amounts in cells of the developing neural tube, but when neural crest cells dissociate from the neural tube and migrate away, they lose N-CAM, only to reexpress it later when they reaggregate to form a neural ganglion. In addition,

Rosenthal et al., (Nature Genet. 2:107-112, 1992) reported that mutations in CAM-L1 (locus Xq28) cause X-linked hydrocephalus, and Jouet et al., (Nature Genet. 7:402-407, 1994) showed that mutations in CAM L1 gene are
5 responsible for type 1 X-linked spastic paraplegia and MASA syndrome which shows agenesis of the corpus callosum. Therefore, there is a need in the art to identify and isolate novel N-CAM members of the immunoglobulin superfamily so that their role in neural
10 development and neural cell communication can be determined.

Therefore, there continues to be a need in the art for the discovery of additional proteins participating in human cell-cell interactions and
15 especially a need for information serving to specifically identify and characterize such proteins in terms of their amino acid sequence. Moreover, to the extent that such molecules might form the basis for the development of therapeutic and diagnostic agents, it is essential that
20 the DNA encoding them be elucidated. The present invention satisfies this need and provides related advantages as well.

BRIEF DESCRIPTION OF THE INVENTION

In accordance with the present invention, there
25 are provided isolated nucleic acids encoding novel mammalian N-CAM (neural-cell adhesion molecule) members of the immunoglobulin superfamily of proteins, referred to herein as Down Syndrome-Cell Adhesion Molecules (DS-CAMs). Further provided are vectors containing
30 invention nucleic acids, probes that hybridize thereto, host cells transformed therewith, antisense oligonucleotides thereto and related compositions. The nucleic acid molecules described herein can be

incorporated into a variety of recombinant expression systems known to those of skill in the art to readily produce isolated DS-CAM proteins. In addition, the nucleic acid molecules of the present invention are
5 useful as probes for assaying for the presence and/or amount of a DS-CAM gene or mRNA transcript in a given sample. The nucleic acid molecules described herein, and oligonucleotide fragments thereof, are also useful as primers and/or templates in a PCR reaction for amplifying
10 genes encoding DS-CAM proteins.

In accordance with the present invention, there are also provided isolated mammalian DS-CAM proteins. These proteins are useful, for example, in neural prosthetic devices used in entubulation methods of
15 repairing (regenerating) damaged or severed peripheral nerves (see, e.g., U.S. Patent No. 4,955,892, incorporated herein by reference). In addition, these proteins, or fragments thereof, are useful as immunogens for producing anti-DS-CAM antibodies, or in therapeutic
20 compositions containing such proteins and/or antibodies. Invention DS-CAM proteins are also useful in bioassays to identify agonists and antagonists thereto. Also provided are transgenic non-human mammals that express the invention protein.

25 Antibodies that are immunoreactive with invention DS-CAM proteins are also provided. These antibodies are useful in diagnostic assays to determine levels of DS-CAM proteins present in a given sample, e.g., tissue samples, Western blots, and the like. The
30 antibodies can also be used to purify DS-CAM proteins from crude cell extracts and the like. Moreover, these antibodies are considered therapeutically useful to counteract or supplement the biological effect of DS-CAMs in vivo.

Methods and diagnostic systems for determining the levels of DS-CAM protein in various tissue samples are also provided. These diagnostic methods can be used for monitoring the level of therapeutically administered DS-CAM protein or fragments thereof to facilitate the maintenance of therapeutically effective amounts. These diagnostic methods can also be used to diagnose physiological disorders that result from abnormal levels or abnormal structures of the DS-CAM protein.

10

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows a physical map of the localization of the DS-CAM gene to a region between D21S345 and D21S347 on chromosome 21. The locations of BAC clones (starting with numbers) and PAC clones (starting with "P") are indicated by horizontal bars. An arrow head indicates a gap in the BAC and PAC contig. The location of the DS-CAM gene is indicated by a thick arrow.

Figure 2 shows the predicted amino acid sequence of the human DS-CAM1 protein corresponding to SEQ ID NO:2 and a schematic structure. **IG:** Immunoglobulin type-C2 domain. **FbN:** Fibronectin type III domain. The bold **Cs** in the amino acid sequence indicates Cysteine residues forming disulfide bonds in the Ig-like type-C2 domains. The bold **NXS** and **NXT** in the amino acid sequence correspond to potential N-glycosylation sites.

Figure 3 shows a partial genomic structure of DS-CAM1 and a deletion contained in DS-CAM2 cDNA clones (clones pDS-CAM-18 and pDS-CAM-52). The deletion boundary sequence (GC-AG) suggests an unusual alternative splicing. The horizontal bar represents

genomic sequence containing exons of DS-CAM-42. Exons are indicated by open boxes. Exon-intron boundaries are defined by a comparison of the cDNA sequence of pDS-CAM-42 and genomic sequence determined from a BAC clone.

Figure 4 shows a schematic comparison of neuronal Ig superfamily members. Ig-like type C-2 domains, fibronectin type III domains and transmembrane domains are indicated. MAG: myelin-associated glycoprotein, N-CAM: neural cell adhesion molecule, BIG-1: brain-derived immunoglobulin (Ig) superfamily molecule-1, DCC: deleted in colorectal carcinoma.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the present invention, there are provided isolated nucleic acids, which encode novel mammalian members of the DS-CAM family of proteins, and fragments thereof. The phrase "DS-CAM" refers to substantially pure native DS-CAM protein, or recombinantly produced proteins, including naturally occurring allelic variants thereof encoded by mRNA generated by alternative splicing of a primary transcript, such as DS-CAM1 (SEQ ID NO:2) and DS-CAM2 (SEQ ID NO:11) disclosed herein, and further including fragments thereof which retain at least one native biological activity, such as immunogenicity. In one aspect, invention DS-CAM proteins, such as DS-CAM1, are cell-surface glycoproteins that are mobile in the plane of the membrane. Invention DS-CAM1 proteins contain extra- and intra-cellular domains that transduce information from the outside of the cell to the cytoplasm and the nucleus, thereby determining cell function. In another aspect, invention DS-CAM proteins, such as DS-CAM2, are non-membrane bound, soluble proteins.

In one aspect of the invention DS-CAM proteins are further characterized as comprising at least 7 Immunoglobulin-like (Ig-like) domains homologous to the immunoglobulin superfamily and 6 type III fibronectin repeats (see, e.g., Edelman and Crossin, "CELL ADHESION MOLECULES: Implications for a Molecular Histology", Ann. Rev. Biochem., **60**:155-190, 1991; and Walsh and Doherty, Curr. Opinion in Cell Biol., **5**:791-796, 1993; each of which is incorporated herein by reference in its entirety). In another aspect of the invention, DS-CAM proteins are those proteins comprising at least 8, preferably at least 9 Ig-like domains, with at least 10 Ig-like domains being especially preferred.

As used herein, "Ig-like domains", or grammatical variations thereof, refers to the well known repeats that are common among Cell Adhesion Molecules (CAMs) (see, e.g., Figure 1A at p. 158 of Edelman and Crossin, supra, 1991; and Walsh and Doherty, supra, 1993; each of which is incorporated herein by reference in its entirety).

The phrase "type III fibronectin repeats", "fibronectin repeats," or grammatical variations thereof, refers to the well known repeats that are common among Cell Adhesion Molecules (CAMs) (see, e.g., Figure 1A at p. 158 of Edelman and Crossin, supra, 1991; and Walsh and Doherty, supra, 1993; each of which is incorporated herein by reference in its entirety).

The invention DS-CAM proteins define a novel sub-class of the Ig (immunoglobulin) superfamily with highest homologies to the neural cell adhesion molecules including BIG-1 (Yoshihara et al., Neuron **13**:415-426, 1994), CAM-L1 (Moos et al., Nature **334**:701-703, 1988), DCC (Fearon et al., Science **247**:49-56, 1990), neogenin

(Lane et al., Genomics **35**:456-465, 1996), and contactin (Ranscht, J. Cell Bio. **107**:1561-1573, 1988) (Figure 4).

It has been found that the structure of invention DS-CAM proteins is unique within the neural immunoglobulin superfamily, and is distinctive due to the number of Ig-like type C2 and fibronectin III domains (10 and 6 respectively) and from the interruption of the fourth and fifth fibronectin domains by a 10th C2 domain, the functional significance of which may be of interest. The novel structure of DS-CAM and its expression throughout the nervous system during differentiation suggest interesting roles for the neural CAM in neural development and function. The location of DS-CAM in a region critical for DS neurocognitive phenotypes provides a human model in which to test the significance of these roles for cognitive function.

The neural Ig-superfamily members play critical roles in neural development and function and have been implicated in cell migration and sorting, axon guidance and fasciculation, formation of neural connections, and in synaptic plasticity (Edelman and Crossin, supra, 1991; Walsh and Doherty, supra, 1993; Tessier-Lavigne et al., Science **274**:1123-1133, 1996; Shuster et al., Neuron **17**:641-654, 1996; Shuster et al., Neuron **17**:655-657, 1996). These activities are mediated by the homophilic or heterophilic binding properties of Ig-superfamily members (Mauro et al., J. Cell Bio. **119**:191-202, 1992 and Milev et al., J. Biol. Chem. **271**:15716-15723, 1996), the binding of Ig-superfamily proteins to extracellular matrix proteins (Grumet et al., Cell Adhesion Comm. **1**:177-190, 1993; Taira et al., Neuron **12** :861-872, 1994; and Zisch et al., J. Cell Bio. **119**:203-213, 1992), and the binding to smaller diffusible chemorepellents or chemoattractants, for example, DCC and netrin (Keino-Masu et al., Cell **87**:175-185, 1996).

The specificity of DS-CAM expression for the central nervous system and the timing of its expression to the period of neurite outgrowth in both the central and peripheral nervous systems, indicates a role for DS-CAM in early development and differentiation (Examples 4 and 5). Early in development when, with the exception of neural crest precursors, expression is clearly absent from regions that contain dividing neuroepithelial precursors such as the ependymal layer of the neural tube and the ventricular zone of the brain (Altman and Bayer, Atlas of Prenatal Rat Brain Development, CRC Press, Ann Arbor, MI, 1995). In the embryo, differentiated neurons express DS-CAM when they have finished migrating to their proper positions within the neuroepithelium, during neurite outgrowth.

Neural crest cells may express DS-CAM while they are migrating. At 15.5 and 16.5 days pc, most of the neural crest derived tissues have some expression, although not all have finished migration. The continued expression of DS-CAM in the myenteric plexus after 15.5-16.5 dpc is due to the neural crest cells that have stopped dividing, although others are in the cell cycle. Approximately 50% of myenteric ganglia neurons arise after birth and DS-CAM may be expressed later in this subset. At later stages, the data suggest that DS-CAM is down regulated in the neural crest derivatives such as the myenteric ganglia and ganglia of the pancreas. The DS-CAM expression in tissues derived from the neural crest is of interest with respect to the high level detected in the umbilical cord. The tissue surrounding the umbilical artery and vein is derived from the neural crest and functions in coordinating the cardiovascular changes occurring at birth. The expression detected in the fetal liver and branchial arches is also derived from neural crest related to the ductus venosus and ultimately

the ductus arteriosus and cardiac outflow tracts,
respectively.

DS-CAM expression continues post-natally, in
the differentiating regions of the newborn brain, such
5 as, the septum and inferior colliculus, and in the adult
in regions associated with plasticity, such as, the
olfactory bulb and hippocampus. When combined with the
evidence for involvement of the Ig superfamily in
determining synaptic strength (Mayford et al., Science
10 **256**:638-644, 1992)), the continued expression supports a
role for DS-CAM in remodeling, learning and memory. The
expression pattern and the role of dendritic connections
in cell body maintenance indicate that an increase in
DS-CAM expression in DS brain is responsible in part for
15 the abnormalities of dendritic structure and decreased
intersections seen at four months post-natal in DS
individuals.

Alternatively spliced variants of CAMs have
distinct roles in different parts of the brain, as
20 demonstrated for closely related Ig-superfamily members,
such as, NCAM (Cunningham et al., Science **236**:799-806,
1987 and Figarella-Branger et al., J. Neuropathol. Exp.
Neurol. **51**:12-23, 1992). The differential expression of
alternatively spliced DS-CAM transcripts encoding DS-CAM1
25 (SEQ ID NO:2) and DS-CAM2 (SEQ ID NO:11) has likewise
been observed in various parts of the human adult brain.
For example, it has been found that DS-CAM clones
encoding DS-CAM2 contain a small deletion relative to
DS-CAM1, which deletion contains the transmembrane domain
30 (Example 3 and Figure 3) and results in a stop codon 36
bp downstream. The results of RT-PCR (Example 5)
indicated that all RNAs tested from various human tissues
expressed both the DS-CAM1 and DS-CAM2 transcripts and
that the PCR products generated the sequence and size
35 predicted for the appropriate form. The proximal and

distal borders of the deletion are located within neighboring exons and reveal variant consensus splice site sequences (Jackson, Nuc. Acid Res. 19:3795-3798, 1991) with further surrounding homology to the U1 spliceosome RNA.

From Northern analyses (Example 4) a minimum of three distinct transcripts are recognized by a probe for the transmembrane domain. From cDNA sequence analyses (Example 5) two forms of the DS-CAM protein are deduced, one that generates a transmembrane adhesion molecule and a second that is deleted for the transmembrane domain, thereby generating a molecule that is transported to the extracellular matrix. This mode of generating extracellular and membrane bound forms of CAMs is in surprising contrast to the GPI (glycosylphosphatidylinositol) linkage used by most CAMs, and would provide a way of generating longer range homophilic interactions between cells and the extracellular matrix, which may be significant for cell migration.

The DS-CAM gene was isolated (as described in the Examples hereinafter) by using the BAC contig on 21q22.2-q22.3 covering the region between D21S55 and MX1 (Hubert et al., Genomics 41:218-226, 1997). The gene spans a minimum of 900 kb, estimated by summing the size of BACs and PACs that are non-overlapping and covered by the DS-CAM gene (Figure 1). The DS-CAM gene covers a gap in all physical maps of this region. From hybridization experiments indicating no signal of the complete cDNA to BAC 277G10 covering 210 kb, a 5' intron is at least this size, similar to the first intron of the DCC gene (Cho et al., Genomics 19:525-531, 1994). Alternatively, other alternative transcripts can contain exons located in this BAC. The gene spans the boundary of bands

21q22.2 and q22.3, a Giemsa-dark and Giemsa-light band, respectively. The location of the gene for PEP19, a small 634 bp gene with large introns within the same band 21q22.2 (Cabin et al., Somat. Cell Mol. Genet. **22**:167-

5 175, 1996) suggests a general structure of genes in G-bands having large introns.

The nucleic acid molecules described herein are useful for producing invention DS-CAM proteins, when such nucleic acids are incorporated into a variety of protein
10 expression systems known to those of skill in the art. In addition, such nucleic acid molecules or fragments thereof can be labeled with a readily detectable substituent and used as hybridization probes for assaying for the presence and/or amount of a DS-CAM gene or mRNA
15 transcript in a given sample. The nucleic acid molecules described herein, and fragments thereof, are also useful as primers and/or templates in a PCR reaction for amplifying genes encoding the invention protein described herein.

20 The term "nucleic acid" (also referred to as polynucleotides) encompasses ribonucleic acid (RNA) or deoxyribonucleic acid (DNA), probes, oligonucleotides, and primers. DNA can be either complementary DNA (cDNA) or genomic DNA, e.g. a gene encoding a DS-CAM protein.
25 One means of isolating a nucleic acid encoding a DS-CAM polypeptide is to probe a mammalian genomic library with a natural or artificially designed DNA probe using methods well known in the art. DNA probes derived from the DS-CAM gene are particularly useful for this purpose.
30 DNA and cDNA molecules that encode DS-CAM polypeptides can be used to obtain complementary genomic DNA, cDNA or RNA from mammalian (e.g., human, mouse, rat, rabbit, pig, and the like), or other animal sources, or to isolate related cDNA or genomic clones by the screening of cDNA
35 or genomic libraries, by methods described in more detail

below. Examples of nucleic acids are RNA, cDNA, or isolated genomic DNA encoding a DS-CAM polypeptide. Such nucleic acids may include, but are not limited to, nucleic acids having substantially the same nucleotide
5 sequence as set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, or at least nucleotides 453-6185 set forth in SEQ ID NO:1, or nucleotides 453-5168 set forth in SEQ ID NO:10.

Use of the terms "isolated" and/or "purified"
10 in the present specification and claims as a modifier of DNA, RNA, polypeptides or proteins means that the DNA, RNA, polypeptides or proteins so designated have been produced in such form by the hand of man, and thus are separated from their native in vivo cellular environment.
15 As a result of this human intervention, the recombinant DNAs, RNAs, polypeptides and proteins of the invention are useful in ways described herein that the DNAs, RNAs, polypeptides or proteins as they naturally occur are not.

As used herein, "mammalian" refers to the
20 variety of species from which the invention DS-CAM protein is derived, e.g., human, rat, mouse, rabbit, monkey, baboon, bovine, porcine, ovine, canine, feline, and the like. A preferred DS-CAM protein herein, is human DS-CAM.

25 In one embodiment of the present invention, cDNAs encoding the invention DS-CAM proteins disclosed herein include substantially the same nucleotide sequence as set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, or SEQ ID NO:10. Preferred cDNA molecules
30 encoding the invention proteins include the same nucleotide sequence as nucleotides 453-6185 set forth in SEQ ID NO:1, or nucleotides 453-5168 set forth in SEQ ID NO:10.

As employed herein, the term "substantially the same nucleotide sequence" refers to DNA having sufficient identity to the reference polynucleotide, such that it will hybridize to the reference nucleotide under

5 moderately stringent hybridization conditions. In one embodiment, DNA having substantially the same nucleotide sequence as the reference nucleotide sequence encodes substantially the same amino acid sequence as that set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM
10 coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9, or a larger amino acid sequence including SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9. In another embodiment, DNA having "substantially the same nucleotide sequence" as
15 the reference nucleotide sequence has at least 60% identity with respect to the reference nucleotide sequence. DNA having at least 70%, more preferably at least 90%, yet more preferably at least 95%, identity to the reference nucleotide sequence is preferred.

20 This invention also encompasses nucleic acids which differ from the nucleic acids shown in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10 but which have the same phenotype. Phenotypically similar nucleic acids are also referred to as "functionally
25 equivalent nucleic acids". As used herein, the phrase "functionally equivalent nucleic acids" encompasses nucleic acids characterized by slight and non-consequential sequence variations that will function in substantially the same manner to produce the same protein
30 product(s) as the nucleic acids disclosed herein. In particular, functionally equivalent nucleic acids encode polypeptides that are the same as those disclosed herein or that have conservative amino acid variations, or that encode larger polypeptides that includes SEQ ID NO:2 or
35 SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9. For example, conservative

variations include substitution of a non-polar residue with another non-polar residue, or substitution of a charged residue with a similarly charged residue. These variations include those recognized by skilled artisans as those that do not substantially alter the tertiary structure of the protein.

Further provided are nucleic acids encoding DS-CAM polypeptides that, by virtue of the degeneracy of the genetic code, do not necessarily hybridize to the invention nucleic acids under specified hybridization conditions. Preferred nucleic acids encoding the invention polypeptides are comprised of nucleotides that encode substantially the same amino acid sequences set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9.

Thus, an exemplary nucleic acid encoding an invention DS-CAM protein may be selected from:

- (a) DNA encoding the amino acid sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9,
- (b) DNA that hybridizes to the DNA of (a) under moderately stringent conditions, wherein said DNA encodes biologically active DS-CAM, or
- (c) DNA degenerate with respect to either (a) or (b) above, wherein said DNA encodes biologically active DS-CAM.

Hybridization refers to the binding of complementary strands of nucleic acid (i.e., sense:antisense strands or probe:target-DNA) to each other through hydrogen bonds, similar to the bonds that naturally occur in chromosomal DNA. Stringency levels used to hybridize a given probe with target-DNA can be readily varied by those of skill in the art.

The phrase "stringent hybridization" is used herein to refer to conditions under which polynucleic acid hybrids are stable. As known to those of skill in the art, the stability of hybrids is reflected in the melting temperature (T_m) of the hybrids. In general, the stability of a hybrid is a function of sodium ion concentration and temperature. Typically, the hybridization reaction is performed under conditions of lower stringency, followed by washes of varying, but higher, stringency. Reference to hybridization stringency relates to such washing conditions.

As used herein, the phrase "moderately stringent hybridization" refers to conditions that permit target-DNA to bind a complementary nucleic acid that has about 60% identity, preferably about 75% identity, more preferably about 85% identity to the target DNA; with greater than about 90% identity to target-DNA being especially preferred. Preferably, moderately stringent conditions are conditions equivalent to hybridization in 50% formamide, 5X Denhardt's solution, 5X SSPE, 0.2% SDS at 42°C, followed by washing in 0.2X SSPE, 0.2% SDS, at 65°C.

The phrase "high stringency hybridization" refers to conditions that permit hybridization of only those nucleic acid sequences that form stable hybrids in 0.018M NaCl at 65°C (i.e., if a hybrid is not stable in 0.018M NaCl at 65°C, it will not be stable under high stringency conditions, as contemplated herein). High stringency conditions can be provided, for example, by hybridization in 50% formamide, 5X Denhardt's solution, 5X SSPE, 0.2% SDS at 42°C, followed by washing in 0.1X SSPE, and 0.1% SDS at 65°C.

The phrase "low stringency hybridization" refers to conditions equivalent to hybridization in 10%

formamide, 5X Denhardt's solution, 6X SSPE, 0.2% SDS at 42°C, followed by washing in 1X SSPE, 0.2% SDS, at 50°C. Denhardt's solution and SSPE (see, e.g., Sambrook et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989) are well known to those of skill in the art as are other suitable hybridization buffers.

As used herein, the term "degenerate" refers to codons that differ in at least one nucleotide from a reference nucleic acid, e.g., SEQ ID NO:1, but encode the same amino acids as the reference nucleic acid. For example, codons specified by the triplets "UCU", "UCC", "UCA", and "UCG" are degenerate with respect to each other since all four of these codons encode the amino acid serine.

Preferred nucleic acids encoding the invention polypeptide(s) hybridize under moderately stringent, preferably high stringency, conditions to substantially the entire sequence, or in certain embodiments substantial portions (i.e., typically at least 15-30 nucleotides) of the nucleic acid sequence set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10.

The invention nucleic acids can be produced by a variety of methods well-known in the art, e.g., the methods described herein, employing PCR amplification using oligonucleotide primers from various regions of SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, and the like.

In accordance with a further embodiment of the present invention, optionally labeled DS-CAM-encoding cDNAs, or fragments thereof, can be employed to probe library(ies) (e.g., cDNA, genomic, and the like) for

additional nucleic acid sequences encoding novel mammalian DS-CAM proteins. As described in Example 3, construction of mammalian cDNA libraries, preferably a human trisomy 21 fetal brain cDNA library, is well-known in the art. Screening of such a cDNA library is initially carried out under low-stringency conditions, which comprise a temperature of less than about 42°C, a formamide concentration of less than about 50%, and a moderate to low salt concentration.

Presently preferred probe-based screening conditions comprise a temperature of about 37°C, a formamide concentration of about 20%, and a salt concentration of about 5X standard saline citrate (SSC; 20X SSC contains 3M sodium chloride, 0.3M sodium citrate, pH 7.0). Such conditions will allow the identification of sequences which have a substantial degree of similarity with the probe sequence, without requiring perfect homology. The phrase "substantial similarity" refers to sequences which share at least 50% homology. Preferably, hybridization conditions will be selected which allow the identification of sequences having at least 70% homology with the probe, while discriminating against sequences which have a lower degree of homology with the probe. As a result, nucleic acids having substantially the same nucleotide sequence as nucleotides 453-6185 set forth in SEQ ID NO:1, or nucleotides 453-5168 set forth in SEQ ID NO:10, SEQ ID NO:7, SEQ ID NO:8, or SEQ ID NO:9 are obtained.

As used herein, a nucleic acid "probe" is single-stranded DNA or RNA, or analogs thereof, that has a sequence of nucleotides that includes at least 14, at least 20, at least 50, at least 100, at least 200, at least 300, at least 400, or at least 500 contiguous bases that are the same as (or the complement of) any contiguous bases set forth in any of SEQ ID NO:1,

SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10. Preferred regions from which to construct probes include 5' and/or 3' coding regions of SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10. In addition,
5 the entire cDNA encoding region of an invention DS-CAM protein, or the entire sequence corresponding to SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10, may be used as a probe. Probes may be labeled by methods well-known in the art, as described hereinafter,
10 and used in various diagnostic kits.

As used herein, the terms "label" and "indicating means" in their various grammatical forms refer to single atoms and molecules that are either directly or indirectly involved in the production of a
15 detectable signal. Any label or indicating means can be linked to invention nucleic acid probes, expressed proteins, polypeptide fragments, or antibody molecules. These atoms or molecules can be used alone or in conjunction with additional reagents. Such labels are
20 themselves well-known in clinical diagnostic chemistry.

The labeling means can be a fluorescent labeling agent that chemically binds to antibodies or antigens without denaturation to form a fluorochrome (dye) that is a useful immunofluorescent tracer. A
25 description of immunofluorescent analytic techniques is found in DeLuca, "Immunofluorescence Analysis", in Antibody As a Tool, Marchalonis et al., eds., John Wiley & Sons, Ltd., pp. 189-231, 1982, which is incorporated herein by reference.

30 In one embodiment, the indicating group is an enzyme, such as horseradish peroxidase (HRP), glucose oxidase, and the like. In another embodiment, radioactive elements are employed labeling agents. The linking of a label to a substrate, i.e., labeling of

nucleic acid probes, antibodies, polypeptides, and proteins, is well known in the art. For instance, an invention antibody can be labeled by metabolic incorporation of radiolabeled amino acids provided in the culture medium. See, for example, Galfre et al., Meth. Enzymol. **73**:3-46, 1981. Conventional means of protein conjugation or coupling by activated functional groups are particularly applicable. See, for example, Aurameas et al., Scand. J. Immunol. **8(7)**:7-23, 1978; Rodwell et al., Biotech. **3**:889-894, 1984; and U.S. Patent No. 4,493,795.

In accordance with another embodiment of the present invention, there are provided isolated mammalian DS-CAM proteins (preferably human), polypeptides, and fragments thereof encoded by invention nucleic acid. Preferably, DS-CAM proteins referred to herein, are those polypeptides specifically recognized by an antibody that also specifically recognizes a DS-CAM protein including the sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9. Invention isolated DS-CAM proteins are free of cellular components and/or contaminants normally associated with a native in vivo environment.

The invention DS-CAM proteins are further characterized as being primarily expressed in fetal brain and not expressed in fetal lung or fetal liver. For example, the results of Northern analysis (described in Example 4) using human fetal tissues showed that 8.5 kb and 7.6 kb transcripts are expressed only in fetal brain and not expressed in fetal lung, fetal liver and fetal kidney. Northern blot analyses of adult tissues revealed differential expression of three alternative transcripts of 9.7 kb, 8.5 kb and 7.6 kb in different substructures of the brain. The 9.7 kb transcript is highly expressed in the substantia nigra, moderately

expressed in the amygdala and hippocampus, and less expressed in the whole brain. A similar pattern is observed by using a PCR product spanning the 191 bp deletion found in DS-CAM-18 and DS-CAM-52. The placenta
 5 shows faint bands, and the sizes are smaller than those in brain. In skeletal muscle, a faint band (6.5 kb) is detected.

The results of RT-PCR (Example 5) demonstrated expression of human DS-CAM mRNA in fetal and adult brain,
 10 in fetal kidney, as well as in a breast carcinoma cell line mRNA. Thus, splice variant cDNA transcripts encoding a DS-CAM family of proteins are clearly contemplated by the present invention.

The region of chromosome locus 21q22.2 from
 15 which DS-CAM is derived is part of the candidate region for holoprosencephaly type I (HPE1). In addition, some patients with this region hemizygotously deleted show abnormalities of the corpus callosum and schizencephaly. Therefore, DS-CAM is contemplated as the gene, which when
 20 defective, deleted or present as a duplication, is responsible for holoprosencephaly, agenesis of the corpus callosum and/or structural defects of the brain. In addition, DS-CAM may also be responsible for several phenotypes of Down Syndrome including mental retardation
 25 as well as, more specifically, the abnormal dendritic structure observed in Down Syndrome. Additional roles for DS-CAM were further evaluated by database homology searches using BLAST X/N and TIGR database analyses. Results of these searches indicate that DS-CAM shows
 30 moderate homology to N-CAM-1 (Cunningham et al., Science, **236**:799-806, 1987) and to DCC (Fearon et al., Science, **247**:49-56, 1990).

Presently preferred DS-CAM proteins of the invention include amino acid sequences that are

substantially the same as the protein sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9, as well as biologically active, modified forms thereof.

5 Those of skill in the art will recognize that numerous residues of the above-described sequences can be substituted with other, chemically, sterically and/or electronically similar residues without substantially altering the biological activity of the resulting
10 receptor species. In addition, larger or smaller polypeptide sequences containing substantially the same sequence as SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9, therein (e.g., splice variants) are contemplated.

15

As employed herein, the term "substantially the same amino acid sequence" refers to amino acid sequences having at least about 50%, preferably at least about 60%, more preferably at least about 70% identity with respect
20 to the reference amino acid sequence, and retaining comparable functional and biological activity characteristic of the protein defined by the reference amino acid sequence. In another embodiment of the invention, preferred invention proteins having
25 "substantially the same amino acid sequence" will have at least about 80%, more preferably 90% amino acid identity with respect to the reference amino acid sequence; with greater than about 95% amino acid sequence identity being especially preferred. It is recognized, however, that
30 polypeptides (or nucleic acids referred to hereinbefore) containing less than the described levels of sequence identity arising as splice variants or that are modified by conservative amino acid substitutions, or by substitution of degenerate codons are also encompassed
35 within the scope of the present invention.

The term "biologically active" or "functional", when used herein as a modifier of invention DS-CAM protein(s), or polypeptide fragment thereof, refers to a polypeptide that exhibits functional characteristics similar to DS-CAM. For example, one biological activity of DS-CAM is the ability to act as an immunogen for the production of polyclonal and monoclonal antibodies that bind specifically to DS-CAM. Thus, an invention nucleic acid encoding DS-CAM will encode a polypeptide specifically recognized by an antibody that also specifically recognizes the DS-CAM protein including the sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9. Such activity may be assayed by any method known to those of skill in the art. For example, a test-polypeptide encoded by a DS-CAM cDNA can be used to produce antibodies, which are then assayed for their ability to bind to the protein including the sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9. If the antibody binds to the test-polypeptide and the protein including the sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9 with substantially the same affinity, then the polypeptide possesses the requisite biological activity.

The invention DS-CAM proteins can be isolated by a variety of methods well-known in the art, e.g., the methods described herein, the recombinant expression systems described herein, precipitation, gel filtration, ion-exchange, reverse-phase and affinity chromatography, and the like. Other well-known methods are described in Deutscher et al., Guide to Protein Purification: Methods in Enzymology 182 (Academic Press, 1990), which is incorporated herein by reference. Alternatively, the isolated polypeptides of the present invention can be

obtained using well-known recombinant methods as described, for example, in Sambrook et al., supra., 1989).

5 An example of the means for preparing the invention polypeptide(s) is to express nucleic acids encoding the DS-CAM in a suitable host cell, such as a bacterial cell, a yeast cell, an amphibian cell (i.e., oocyte), or a mammalian cell, using methods well known in the art, and recovering the expressed polypeptide, again
10 using well-known methods. Invention polypeptides can be isolated directly from cells that have been transformed with expression vectors as described below herein. The invention polypeptide, biologically active fragments, and functional equivalents thereof can also be produced by
15 chemical synthesis. For example, synthetic polypeptides can be produced using Applied Biosystems, Inc. Model 430A or 431A automatic peptide synthesizer (Foster City, CA) employing the chemistry provided by the manufacturer.

The present invention also provides
20 compositions containing an acceptable carrier and any of an isolated, purified DS-CAM polypeptide, an active fragment thereof, or a purified, mature protein and active fragments thereof, alone or in combination with each other. These polypeptides or proteins can be
25 recombinantly derived, chemically synthesized or purified from native sources. As used herein, the term "acceptable carrier" encompasses any of the standard pharmaceutical carriers, such as phosphate buffered saline solution, water and emulsions such as an oil/water
30 or water/oil emulsion, and various types of wetting agents.

Also provided are antisense oligonucleotides having a sequence capable of binding specifically with any portion of an mRNA that encodes DS-CAM polypeptides

so as to prevent translation of the mRNA. The antisense oligonucleotide may have a sequence capable of binding specifically with any portion of the sequence of the cDNA encoding DS-CAM polypeptides. As used herein, the phrase
5 "binding specifically" encompasses the ability of a nucleic acid sequence to recognize a complementary nucleic acid sequence and to form double-helical segments therewith via the formation of hydrogen bonds between the complementary base pairs. An example of an antisense
10 oligonucleotide is an antisense oligonucleotide comprising chemical analogs of nucleotides.

Compositions comprising an amount of the antisense oligonucleotide, described above, effective to reduce expression of DS-CAM polypeptides by passing
15 through a cell membrane and binding specifically with mRNA encoding DS-CAM polypeptides so as to prevent translation and an acceptable hydrophobic carrier capable of passing through a cell membrane are also provided herein. Suitable hydrophobic carriers are described, for
20 example, in U.S. Patent Nos. 5,334,761; 4,889,953; 4,897,355, and the like. The acceptable hydrophobic carrier capable of passing through cell membranes may also comprise a structure which binds to a receptor specific for a selected cell type and is thereby taken up
25 by cells of the selected cell type. The structure may be part of a protein known to bind to a cell-type specific receptor.

Antisense oligonucleotide compositions are useful to inhibit translation of mRNA encoding invention
30 polypeptides. Synthetic oligonucleotides, or other antisense chemical structures are designed to bind to mRNA encoding DS-CAM polypeptides and inhibit translation of mRNA and are useful as compositions to inhibit expression of DS-CAM associated genes in a tissue sample
35 or in a subject.

In accordance with another embodiment of the invention, kits for detecting mutations, duplications, deletions, rearrangements and aneuploidies in chromosome 21 at locus q22.2 comprising at least one invention probe
5 or antisense nucleotide.

The present invention provides means to modulate levels of expression of DS-CAM polypeptides by employing synthetic antisense oligonucleotide compositions (hereinafter SAOC) which inhibit translation
10 of mRNA encoding these polypeptides. Synthetic oligonucleotides, or other antisense chemical structures designed to recognize and selectively bind to mRNA, are constructed to be complementary to portions of the DS-CAM coding strand or nucleotide sequences shown in
15 SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10. The SAOC is designed to be stable in the blood stream for administration to a subject by injection, or in laboratory cell culture conditions. The SAOC is designed to be capable of passing through the
20 cell membrane in order to enter the cytoplasm of the cell by virtue of physical and chemical properties of the SAOC which render it capable of passing through cell membranes, for example, by designing small, hydrophobic SAOC chemical structures, or by virtue of specific
25 transport systems in the cell which recognize and transport the SAOC into the cell. In addition, the SAOC can be designed for administration only to certain selected cell populations by targeting the SAOC to be recognized by specific cellular uptake mechanisms which
30 bind and take up the SAOC only within select cell populations.

For example, the SAOC may be designed to bind to a receptor found only in a certain cell type, as discussed supra. The SAOC is also designed to recognize
35 and selectively bind to target mRNA sequence, which may

correspond to a sequence contained within the sequence shown in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8,

SEQ ID NO:9 or SEQ ID NO:10. The SAOC is designed to inactivate target mRNA sequence by either binding thereto
5 and inducing degradation of the mRNA by, for example, RNase I digestion, or inhibiting translation of mRNA target sequence by interfering with the binding of translation-regulating factors or ribosomes, or inclusion
10 of other chemical structures, such as ribozyme sequences or reactive chemical groups which either degrade or chemically modify the target mRNA. SAOCs have been shown to be capable of such properties when directed against mRNA targets (see Cohen et al., TIBS 10:435, 1989 and Weintraub, Sci. American January 1990, pp.40; both
15 incorporated herein by reference).

In accordance with yet another embodiment of the present invention, there is provided a method for the recombinant production of invention DS-CAM protein(s) by expressing the above-described nucleic acid sequences in
20 suitable host cells. Recombinant DNA expression systems that are suitable to produce DS-CAM proteins described herein are well-known in the art. For example, the above-described nucleotide sequences can be incorporated into vectors for further manipulation. As used herein,
25 vector (or plasmid) refers to discrete elements that are used to introduce heterologous DNA into cells for either expression or replication thereof.

Suitable expression vectors are well-known in the art, and include vectors capable of expressing DNA
30 operatively linked to a regulatory sequence, such as a promoter region that is capable of regulating expression of such DNA. Thus, an expression vector refers to a recombinant DNA or RNA construct, such as a plasmid, a phage, recombinant virus or other vector that, upon
35 introduction into an appropriate host cell, results in

expression of the inserted DNA. Appropriate expression vectors are well known to those of skill in the art and include those that are replicable in eukaryotic cells and/or prokaryotic cells and those that remain episomal or those which integrate into the host cell genome.

As used herein, a promoter region refers to a segment of DNA that controls transcription of DNA to which it is operatively linked. The promoter region includes specific sequences that are sufficient for RNA polymerase recognition, binding and transcription initiation. In addition, the promoter region includes sequences that modulate this recognition, binding and transcription initiation activity of RNA polymerase. These sequences may be *cis* acting or may be responsive to *trans* acting factors. Promoters, depending upon the nature of the regulation, may be constitutive or regulated. Exemplary promoters contemplated for use in the practice of the present invention include the SV40 early promoter, the cytomegalovirus (CMV) promoter, the mouse mammary tumor virus (MMTV) steroid-inducible promoter, Moloney murine leukemia virus (MMLV) promoter, and the like.

As used herein, the term "operatively linked" refers to the functional relationship of DNA with regulatory and effector nucleotide sequences, such as promoters, enhancers, transcriptional and translational stop sites, and other signal sequences. For example, operative linkage of DNA to a promoter refers to the physical and functional relationship between the DNA and the promoter such that the transcription of such DNA is initiated from the promoter by an RNA polymerase that specifically recognizes, binds to and transcribes the DNA.

As used herein, expression refers to the process well-known to those of skill in the art by which polynucleic acids are transcribed into mRNA and translated into peptides or proteins and, optionally thereafter, modified post-translationally. If the invention nucleic acid is derived from genomic DNA, expression may, if an appropriate eukaryotic host cell or organism is selected, include splicing of the mRNA.

Prokaryotic transformation vectors are well-known in the art and include pBluescript and phage Lambda ZAP vectors (STRATAGENE, San Diego, CA), and the like. Other suitable vectors and promoters are disclosed in detail in U.S. Patent No. 4,798,885, issued January 17, 1989, the disclosure of which is incorporated herein by reference in its entirety.

Other suitable vectors for transformation of *E. coli* cells include the pET expression vectors (Novagen, see U.S patent 4,952,496), e.g., pET11a, which contains the T7 promoter, T7 terminator, the inducible *E. coli* lac operator, and the lac repressor gene; and pET 12a-c, which contain the T7 promoter, T7 terminator, and the *E. coli* ompT secretion signal. Another suitable vector is the pIN-IIIompA2 (see Duffaud et al., Meth. in Enzymology, **153**:492-507, 1987), which contains the lpp promoter, the lacUV5 promoter operator, the ompA secretion signal, and the lac repressor gene.

Exemplary, eukaryotic transformation vectors, include the cloned bovine papilloma virus genome, the cloned genomes of the murine retroviruses, and eukaryotic cassettes, such as the pSV-2 gpt system (described by Mulligan and Berg, Nature **277**:108-114, 1979) the Okayama-Berg cloning system (Mol. Cell Biol. **2**:161-170, 1982), and the expression cloning vector described by

Genetics Institute (Science 228:810-815, 1985), are available which provide substantial assurance of at least some expression of the protein of interest in the transformed eukaryotic cell line.

5 Particularly preferred base vectors which contain regulatory elements that can be linked to the invention DS-CAM-encoding DNAs for transfection of mammalian cells are cytomegalovirus (CMV) promoter-based vectors such as pcDNA1 (Invitrogen, San Diego, CA), MMTV
10 promoter-based vectors such as pMAMNeo (Clontech, Palo Alto, CA) and pMSG (Pharmacia, Piscataway, NJ), and SV40 promoter-based vectors such as pSV β (Clontech, Palo Alto, CA).

15 In accordance with another embodiment of the present invention, there are provided "recombinant cells" containing the nucleic acid molecules (i.e., DNA or mRNA) of the present invention. Methods of transforming suitable host cells, preferably bacterial cells, and more preferably *E. coli* cells, as well as methods applicable
20 for culturing said cells containing a gene encoding a heterologous protein, are generally known in the art. See, for example, Sambrook et al., supra, 1989.

25 Exemplary methods of transformation include, e.g., transformation employing plasmids, viral, or bacterial phage vectors, transfection, electroporation, lipofection, and the like. The heterologous DNA can optionally include sequences which allow for its extrachromosomal maintenance, or said heterologous DNA can be caused to integrate into the genome of the host
30 (as an alternative means to ensure stable maintenance in the host).

Host organisms contemplated for use in the practice of the present invention include those organisms

in which recombinant production of heterologous proteins has been carried out. Exemplary cells for introducing DNA include cells of mammalian origin (e.g., COS cells, mouse L cells, Chinese hamster ovary (CHO) cells, human
5 embryonic kidney (HEK) cells, African green monkey cells and other such cells known to those of skill in the art), amphibian cells (e.g., *Xenopus laevis* oöcytes), yeast cells (e.g., *Saccharomyces cerevisiae*, *Candida tropicalis*, *Hansenula polymorpha* and *P. pastoris*; see,
10 e.g., U.S. Patent Nos. 4,882,279, 4,837,148, 4,929,555 and 4,855,231), bacteria (e.g., *E. coli*), and the like.

In one embodiment, nucleic acids encoding the invention DS-CAM proteins can be delivered into mammalian cells, either in vivo or in vitro using suitable viral
15 vectors well-known in the art. Suitable retroviral vectors, designed specifically for in vivo "gene therapy" methods, are described, for example, in WIPO publications WO 9205266 and WO 9214829, which provide a description of methods for efficiently introducing nucleic acids into
20 human cells in vivo. In addition, where it is desirable to limit or reduce the in vivo expression of the invention DS-CAM, the introduction of the antisense strand of the invention nucleic acid is contemplated.

In accordance with yet another embodiment of
25 the present invention, there are provided anti-DS-CAM antibodies having specific reactivity with DS-CAM polypeptides of the present invention. Active fragments of antibodies are encompassed within the definition of "antibody". Invention antibodies can be produced by
30 methods known in the art using invention polypeptides, proteins or portions thereof as antigens. For example, polyclonal and monoclonal antibodies can be produced by methods well known in the art, as described, for example, in Harlow and Lane, Antibodies: A Laboratory Manual (Cold

Spring Harbor Laboratory, 1988), which is incorporated herein by reference. Invention polypeptides can be used as immunogens in generating such antibodies. Alternatively, synthetic peptides can be prepared (using
5 commercially available synthesizers) and used as immunogens. Amino acid sequences can be analyzed by methods well known in the art to determine whether they encode hydrophobic or hydrophilic domains of the corresponding polypeptide. Altered antibodies such as
10 chimeric, humanized, CDR-grafted or bifunctional antibodies can also be produced by methods well known in the art. Such antibodies can also be produced by hybridoma, chemical synthesis or recombinant methods described, for example, in Sambrook et al., supra, 1989; and Harlow and Lane, supra, 1988. Both anti-peptide and anti-fusion protein antibodies can be used. (see, for example, Bahouth et al., Trends Pharmacol. Sci. 12:338
15 1991; Ausubel et al., Current Protocols in Molecular Biology (John Wiley and Sons, NY 1989) which are
20 incorporated herein by reference).

Antibody so produced can be used, inter alia, in diagnostic methods and systems to detect the level of DS-CAM protein present in a mammalian, preferably human, body sample, such as tissue or vascular fluid. Such
25 antibodies can also be used for the immunoaffinity or affinity chromatography purification of the invention DS-CAM protein. In addition, methods are contemplated herein for detecting the presence of DS-CAM polypeptides on the surface of a cell comprising contacting the cell
30 with an antibody that specifically binds to DS-CAM polypeptides, under conditions permitting binding of the antibody to the polypeptides, detecting the presence of the antibody bound to the cell, and thereby detecting the presence of invention polypeptides on the surface of the
35 cell. With respect to the detection of such

polypeptides, the antibodies can be used for in vitro diagnostic or in vivo imaging methods.

Immunological procedures useful for in vitro detection of target DS-CAM polypeptides in a sample
5 include immunoassays that employ a detectable antibody. Such immunoassays include, for example, ELISA, Pandex microfluorimetric assay, agglutination assays, flow cytometry, serum diagnostic assays and immunohistochemical staining procedures which are well
10 known in the art. An antibody can be made detectable by various means well known in the art. For example, a detectable marker can be directly or indirectly attached to the antibody. Useful markers include, for example, radionucleotides, enzymes, fluorogens, chromogens and
15 chemiluminescent labels.

Invention anti-DS-CAM antibodies are contemplated for use herein to modulate the activity of the DS-CAM polypeptide in living animals, in humans, or in biological tissues or fluids isolated therefrom.
20 Accordingly, compositions comprising a carrier and an amount of an antibody having specificity for DS-CAM polypeptides effective to block naturally occurring ligands or other DS-CAM-binding proteins from binding to invention DS-CAM polypeptides are contemplated herein.
25 For example, a monoclonal antibody directed to an epitope of DS-CAM polypeptide molecules present on the surface of a cell and having an amino acid sequence substantially the same as an amino acid sequence for a cell surface epitope of a DS-CAM polypeptide including the amino acid
30 sequence shown in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9, can be useful for this purpose.

The present invention further provides transgenic non-human mammals that are capable of

expressing exogenous nucleic acids encoding DS-CAM polypeptides. As employed herein, the phrase "exogenous nucleic acid" refers to nucleic acid sequence which is not native to the host, or which is present in the host in other than its native environment (e.g., as part of a genetically engineered DNA construct).

Also provided are transgenic non-human mammals capable of expressing nucleic acids encoding DS-CAM polypeptides so mutated as to be incapable of normal activity, i.e., do not express native DS-CAM. The present invention also provides transgenic non-human mammals having a genome comprising antisense nucleic acids complementary to nucleic acids encoding DS-CAM polypeptides, placed so as to be transcribed into antisense mRNA complementary to mRNA encoding DS-CAM polypeptides, which hybridizes to the mRNA and, thereby, reduces the translation thereof. The nucleic acid may additionally comprise an inducible promoter and/or tissue specific regulatory elements, so that expression can be induced, or restricted to specific cell types. Examples of nucleic acids are DNA or cDNA having a coding sequence substantially the same as the coding sequence shown in SEQ ID NO:1. An example of a non-human transgenic mammal is a transgenic mouse. Examples of tissue specificity-determining elements are the metallothionein promoter and the L7 promoter.

Animal model systems which elucidate the physiological and behavioral roles of DS-CAM polypeptides are also provided, and are produced by creating transgenic animals in which the expression of the DS-CAM polypeptide is altered using a variety of techniques. Examples of such techniques include the insertion of normal or mutant versions of nucleic acids encoding a DS-CAM polypeptide by microinjection, retroviral infection or other means well known to those skilled in

the art, into appropriate fertilized embryos to produce a transgenic animal. See, for example, Hogan et al., Manipulating the Mouse Embryo: A Laboratory Manual (Cold Spring Harbor Laboratory, 1986).

5 Also contemplated herein, is the use of homologous recombination of mutant or normal versions of DS-CAM genes with the native gene locus in transgenic animals, to alter the regulation of expression or the structure of DS-CAM polypeptides (see, Capecchi et al.,
10 Science **244**:1288, 1989; Zimmer et al., Nature **338**:150, 1989; which are incorporated herein by reference). Homologous recombination techniques are well known in the art. Homologous recombination replaces the native (endogenous) gene with a recombinant or mutated gene to
15 produce an animal that cannot express native (endogenous) protein but can express, for example, a mutated protein which results in altered expression of DS-CAM polypeptides.

 In contrast to homologous recombination,
20 microinjection adds genes to the host genome, without removing host genes. Microinjection can produce a transgenic animal that is capable of expressing both endogenous and exogenous DS-CAM protein. Inducible promoters can be linked to the coding region of nucleic
25 acids to provide a means to regulate expression of the transgene. Tissue specific regulatory elements can be linked to the coding region to permit tissue-specific expression of the transgene. Transgenic animal model systems are useful for in vivo screening of compounds for
30 identification of specific ligands, i.e., agonists and antagonists, which activate or inhibit protein responses.

 Invention nucleic acids, oligonucleotides (including antisense), vectors containing same, transformed host cells, polypeptides and combinations

thereof, as well as antibodies of the present invention, can be used to screen compounds in vitro to determine whether a compound functions as a potential agonist or antagonist to invention polypeptides. These in vitro screening assays provide information regarding the function and activity of invention polypeptides, which can lead to the identification and design of compounds that are capable of specific interaction with one or more types of polypeptides, peptides or proteins.

10 In accordance with still another embodiment of the present invention, there is provided a method for identifying compounds which bind to DS-CAM polypeptides. The invention proteins may be employed in a competitive binding assay. Such an assay can accommodate the rapid screening of a large number of compounds to determine which compounds, if any, are capable of binding to DS-CAM proteins. Subsequently, more detailed assays can be carried out with those compounds found to bind, to further determine whether such compounds act as modulators, agonists or antagonists of invention proteins.

Another application of the binding assay of the invention is the assay of test samples (e.g., biological fluids) for the presence or absence of DS-CAM. Thus, for example, serum from a patient displaying symptoms thought to be related to over- or under-production of DS-CAM can be assayed to determine if the observed symptoms are indeed caused by over- or under-production of DS-CAM.

In another embodiment of the invention, there is provided a bioassay for identifying compounds which modulate the activity of invention DS-CAM polypeptides. According to this method, invention polypeptides are contacted with an "unknown" or test substance (in the presence of a reporter gene construct when antagonist

activity is tested), the activity of the polypeptide is monitored subsequent to the contact with the "unknown" or test substance, and those substances which cause the reporter gene construct to be expressed are identified as functional ligands for DS-CAM polypeptides.

In accordance with another embodiment of the present invention, transformed host cells that recombinantly express invention polypeptides can be contacted with a test compound, and the modulating effect(s) thereof can then be evaluated by comparing the DS-CAM-mediated response (e.g., via reporter gene expression) in the presence and absence of test compound, or by comparing the response of test cells or control cells (i.e., cells that do not express DS-CAM polypeptides), to the presence of the compound.

As used herein, a compound or a signal that "modulates the activity" of invention polypeptides refers to a compound or a signal that alters the activity of DS-CAM polypeptides so that the activity of the invention polypeptide is different in the presence of the compound or signal than in the absence of the compound or signal. In particular, such compounds or signals include agonists and antagonists. An agonist encompasses a compound or a signal that activates DS-CAM protein expression. Alternatively, an antagonist includes a compound or signal that interferes with DS-CAM protein expression. Typically, the effect of an antagonist is observed as a blocking of agonist-induced protein activation. Antagonists include competitive and non-competitive antagonists. A competitive antagonist (or competitive blocker) interacts with or near the site specific for agonist binding. A non-competitive antagonist or blocker inactivates the function of the polypeptide by interacting with a site other than the agonist interaction site.

As understood by those of skill in the art, assay methods for identifying compounds that modulate DS-CAM activity generally require comparison to a control. One type of a "control" is a cell or culture
5 that is treated substantially the same as the test cell or test culture exposed to the compound, with the distinction that the "control" cell or culture is not exposed to the compound. For example, in methods that use voltage clamp electrophysiological procedures, the
10 same cell can be tested in the presence or absence of compound, by merely changing the external solution bathing the cell. Another type of "control" cell or culture may be a cell or culture that is identical to the transfected cells, with the exception that the "control"
15 cell or culture do not express native proteins. Accordingly, the response of the transfected cell to compound is compared to the response (or lack thereof) of the "control" cell or culture to the same compound under the same reaction conditions.

20 Since it is well-known that CAMs interact with extracellular ligands, it is contemplated that invention DS-CAM proteins interact with extracellular ligands. In another embodiment of the present invention, it is contemplated that invention DS-CAM proteins act
25 specifically in concert or in competition with other CAMs. Thus, the present invention contemplates various bioassays for identifying ligands for invention DS-CAM proteins. In addition, the present invention contemplates an assay measuring the effect of
30 co-expressing during development either normal or defective invention DS-CAMs with other CAMs known in the art to assess the resulting phenotype.

In one embodiment of the present invention,
35 there is provided a bioassay for evaluating whether test compounds are capable of acting as agonists comprises:

(a) culturing cells containing:

DNA which expresses DS-CAM
protein(s) or functional modified
forms thereof, and

5 DNA encoding a reporter protein,
wherein said DNA is operatively
linked to a DS-CAM responsive
transcription element;

10 wherein said culturing is carried out in
the presence of at least one compound
whose ability to induce signal
transduction activity of DS-CAM protein is
sought to be determined, and thereafter

15 (b) monitoring said cells for expression of
said reporter protein.

In another embodiment of the present invention,
the bioassay for evaluating whether test compounds are
capable of acting as antagonists for DS-CAM protein(s) of
the invention, or functional modified forms of said
20 DS-CAM protein(s), comprises:

(a) culturing cells containing:

DNA which expresses DS-CAM
protein(s), or functional modified
forms thereof, and

25 DNA encoding a reporter protein,
wherein said DNA is operatively
linked to a DS-CAM responsive
transcription element

30 wherein said culturing is carried out in
the presence of:

35 increasing concentrations of at
least one compound whose ability to
inhibit signal transduction activity
of DS-CAM protein(s) is sought to be
determined, and

a fixed concentration of at least one agonist for DS-CAM protein(s), or functional modified forms thereof; and thereafter

- 5 (b) monitoring in said cells the level of expression of said reporter protein as a function of the concentration of said compound, thereby indicating the ability of said compound to inhibit signal
10 transduction activity.

In step (a) of the above-described antagonist bioassay, culturing may also be carried out in the presence of:

- 15 fixed concentrations of at least one compound whose ability to inhibit signal transduction activity of DS-CAM protein(s) is sought to be determined, and
an increasing concentration of at least one agonist for DS-CAM
20 protein(s), or functional modified forms thereof.

In yet another embodiment of the present invention, it is contemplated that invention DS-CAM proteins mediate signal transduction through the
25 modulation of adenylate cyclase. For example, when a DS-CAM ligand binds to DS-CAM, adenylate cyclase causes an elevation in the level of intracellular cAMP. Accordingly, in one embodiment of the present invention, the bioassay for evaluating whether test compounds are
30 capable of acting as agonists or antagonists comprises:

- (a) culturing cells containing:

DNA which expresses DS-CAM protein(s) or functional modified forms thereof,

wherein said culturing is carried out in the presence of at least one compound whose ability to modulate signal transduction activity of DS-CAM protein is sought to be determined, and thereafter

5 (b) monitoring said cells for either an increase or decrease in the level of intracellular cAMP.

Methods well-known in the art that measure

10 intracellular levels of cAMP, or measure cyclase activity, can be employed in binding assays described herein to identify agonists and antagonists of the DS-CAM. For example, because activation of some CAMs results in decreases or increases in cAMP, assays that

15 measure intracellular cAMP levels can be used to evaluate recombinant DS-CAMs expressed in mammalian host cells.

As used herein, "ability to modulate signal transduction activity of DS-CAM protein" refers to a compound that has the ability to either induce (agonist)

20 or inhibit (antagonist) signal transduction activity of the DS-CAM protein.

Each of the invention bioassays (e.g., those described herein, and the like), can be conducted as competitive assays by co-expressing one or more members

25 of the CAM immunoglobulin superfamily of proteins known in the art, such as N-CAMs, along with invention DS-CAMs. In addition, one or more members of the CAM immunoglobulin superfamily of proteins known in the art can be co-expressed with invention DS-CAMs to evaluate

30 the agonistic or antagonistic effect on signal transduction of the non-DS-CAM members acting in concert with invention DS-CAMs.

In yet another embodiment of the present invention, the activation of DS-CAM polypeptides can be modulated by contacting the polypeptides with an effective amount of at least one compound identified by the above-described bioassays.

Members of the N-CAM superfamily of immunoglobulins have previously been implicated in disease. For example, various alterations of N-CAM levels have been seen in degenerative disease, developmental defects, and toxic conditions. Increases in the levels of N-CAM in the cerebrospinal fluid of patients with multiple sclerosis have been observed to parallel their clinical improvement (Massaro et al., Ital. J. Neurol. Sci. Suppl. 6:85-88, 1987). Levels of N-CAM were reported to be elevated in the amniotic fluid of mothers carrying fetuses with neural tube defects (Ibsen et al., J. Neurochem. 41:363-366, 1983). Since many such defects are likely to be due to mechanical aberrations rather than genetic defects, confirmation of these results would provide a new diagnostic component for prenatal testing. Another provocative finding relates to observations on the stimulation of Golgi sialyltransferases by lead (Breen and Regan, Development 104:147-154, 1988; and Cookman et al., J. Neurochem. 49:399-403, 1987). Exposure to lead chloride markedly stimulated sialyltransferase activity from postnatal days 16 to 30 in rate. This time is coincident with the period when N-CAM normally becomes less sialylated. Thus exposure to lead at critical developmental periods would presumably lead to more highly sialylated, less adhesive, forms of N-CAM: this prevention of E-A conversion could have significant effects on neural development. E-A conversion itself has been found to be delayed in the mouse mutant *staggerer* (Edelman and Chuong, Proc. Natl.

Acad. Sci. USA, **79**:7036-7042, 1982) in conjunction with the connectivity changes associated with the mutation.

The location and expression of DS-CAM in the Down Syndrome (DS) phenotype is supported by the studies of patients with partial trisomy 21. A subset of the DS features, including the typical facial appearance and mental retardation, were suggested by duplication of band 21q22 only (Niebuhr, Humangenetik **21**:99-101, 1974).

Other studies mapped those features and congenital heart disease to the region 21q22.2-q22.3 and between D21S267 and MX1/MX2 (Korenberg et al., Am. J. Hum. Genet. **50**:294-302, 1992 and Korenberg et al., Proc. Natl. Acad. Sci. USA **91**:4997-5001, 1994), a region of about 4 Mb that contains DS-CAM. The Ts65Dn mouse model of DS contains the region of MMU16 (Pgk1-ps1 to MX1/2) that includes DS-CAM and reveals some of the neurobehavioural features of DS (Reeves et al., Nature Genet. **11**:177-183, 1995 and Holtzman et al., Proc. Natl. Acad. Sci. USA **93**:13333-13338, 1996).

Close to 6% of DS individuals have Hirschsprung's disease (HSCR) (Garver et al., Clin. Genet. **28**:503-5-8, 1985) and more than 10% of all HSCR is associated with DS (Passarge, New Eng. J. Med. **276**:138-143, 1967). A modifier region of HSCR on chromosome 21q22 (D21S259 - D21S156) has been reported in non-DS HSCR (Puffenberger et al., Hum. Mol. Genet. **3**:1217-1225, 1994). The DS-CAM gene maps within this small region. The expression of DS-CAM in the neural crest derived enteric plexus of the gut was detected by mouse tissue in situ hybridization (Example 7). The function of the DS-CAM protein as a neural cell adhesion molecule and the association of this region of chromosome 21 with HSCR, indicate that DS-CAM can play a role in the migration of

the cranial neural crest that populate this region. Thus, DS-CAM overexpression is responsible for the chromosome 21 association in non-DS HSCR and for the HSCR seen in DS.

5 Mutations in the molecule CAM-L1, a molecule more similar to DS-CAM than to N-CAM (Figure 4), have established roles in human disease. The result in X-linked hydrocephalus (Rosenthal et al., Nature Genet. 2:107-112, 1992), type 1 X-linked spastic paraplegia and
10 the MASA syndrome (including mental retardation, aphasia, shuffling gait, adducted thumb and agenesis of the corpus callosum) (Jouet et al., Nature Genet. 7:402-407, 1994). The perturbation of development by the aneuploid expression of CAM-L1 supports a role for the aneuploid
15 expression of DS-CAM in the causation of developmental and neurological abnormalities.

In accordance with another embodiment of the present invention, there are provided methods for diagnosing DS-CAM associated disease, such as mental
20 retardation, holoprosencephaly, agenesis of the corpus callosum, or schizencephaly, said method comprising:
detecting, in said subject, a genomic or transcribed mRNA sequence including SEQ ID NO:1 or SEQ ID NO:10, or fragments thereof.

25 Preferably, the DS-CAM nucleic acids detected in accordance with the invention diagnostic methods are either mutated in one form or another (such as point mutations, deletions, and the like), or are overexpressed relative to levels of DS-CAM expression in healthy
30 non-diseased individuals.

In accordance with another embodiment of the present invention, there are provided diagnostic systems,

preferably in kit form, comprising at least one invention nucleic acid in a suitable packaging material. The diagnostic nucleic acids are derived from the DS-CAM-encoding nucleic acids described herein. In one
5 embodiment, for example, the diagnostic nucleic acids are derived from SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10. Invention diagnostic systems are useful for assaying for the presence or absence of nucleic acid encoding DS-CAM in either genomic
10 DNA or in transcribed nucleic acid (such as mRNA or cDNA) encoding DS-CAM.

A suitable diagnostic system includes at least one invention nucleic acid, preferably two or more invention nucleic acids, as a separately packaged
15 chemical reagent(s) in an amount sufficient for at least one assay. Instructions for use of the packaged reagent are also typically included. Those of skill in the art can readily incorporate invention nucleic probes and/or primers into kit form in combination with appropriate
20 buffers and solutions for the practice of the invention methods as described herein.

As employed herein, the phrase "packaging material" refers to one or more physical structures used to house the contents of the kit, such as invention
25 nucleic acid probes or primers, and the like. The packaging material is constructed by well known methods, preferably to provide a sterile, contaminant-free environment. The packaging material has a label which indicates that the invention nucleic acids can be used
30 for detecting a particular sequence encoding DS-CAM including the nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10, thereby diagnosing the presence of, or a predisposition for, holoprosencephaly, agenesis of the
35 corpus callosum, or for several phenotypes of Down

Syndrome including mental retardation, and the like. In addition, the packaging material contains instructions indicating how the materials within the kit are employed both to detect a particular sequence and diagnose the
5 presence of, or a predisposition for, holoprosencephaly, agenesis of the corpus callosum, or for several phenotypes of Down syndrome including mental retardation, and the like.

The packaging materials employed herein in
10 relation to diagnostic systems are those customarily utilized in nucleic acid-based diagnostic systems. As used herein, the term "package" refers to a solid matrix or material such as glass, plastic, paper, foil, and the like, capable of holding within fixed limits an isolated
15 nucleic acid, oligonucleotide, or primer of the present invention. Thus, for example, a package can be a glass vial used to contain milligram quantities of a contemplated nucleic acid, oligonucleotide or primer, or it can be a microtiter plate well to which microgram
20 quantities of a contemplated nucleic acid probe have been operatively affixed.

"Instructions for use" typically include a tangible expression describing the reagent concentration or at least one assay method parameter, such as the
25 relative amounts of reagent and sample to be admixed, maintenance time periods for reagent/sample admixtures, temperature, buffer conditions, and the like.

All U.S. patents and all publications mentioned herein are incorporated in their entirety by reference
30 thereto. The invention will now be described in greater detail by reference to the following non-limiting examples.

Materials and Methods

Unless otherwise stated, the present invention was performed using standard procedures, as described, for example in Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA, 1982; Sambrook et al., *supra*, 1989; Davis et al., Basic Methods in Molecular Biology, Elsevier Science Publishing, Inc., New York, USA, 1986; or Methods in Enzymology: Guide to Molecular Cloning Techniques Vol. 152, S. L. Berger and A. R. Kimmerl Eds., Academic Press Inc., San Diego, USA, 1987.

Libraries.

Construction of Bacterial Artificial Chromosome

(BAC) library. BAC library construction of total human genomic DNA was performed as described in Shizuya et al., Proc. Natl. Acad. Sci. USA **89**:8794-8797, 1992; and Hubert et al., Genomics **41**:218-226, 1997. Yeast artificial chromosome (YAC) clones were obtained from the CEPH mega-YAC library and grown under standard conditions (Cohen et al., Nature **366**:689-701 1993).

P1 artificial chromosome (PAC) library

construction. A 3X human PAC library, designated RPCI-1 (Ioannou et al., Hum. Genet. 219-220, 1994) was constructed as described (Ioannou et al., Nat. Genet. **6**:84-89, 1994). The library was arrayed in 384 well dishes. Subsequently, STSs generated by sequencing of clones using vector primers were used as hybridization probes to gridded colony filters of the PAC library.

YAC DNA preparation. YAC clones were grown in selective media, pelleted and resuspended in 3 ml 0.9 M sorbitol, 0.1M EDTA pH 7.5, then incubated with 100 U of

lytcase (Sigma, St. Louis, MO) at 37°C for 1 hour. After centrifugation for 5 minutes at 5,000 rpm pellets were resuspended in 3 ml 50 mM Tris pH 7.45, 20 mM EDTA 0.3ml 10% SDS was added and the mixture was incubated at 65°C
 5 for 30 minutes. One ml of 5 M potassium acetate was added and tubes were left on ice for 1 hour, then centrifuged at 10,000 rpm for 10 minutes. Supernatant was precipitated in 2 volumes of ethanol and pelleted at 6,000 rpm for 15 minutes. Pellets were resuspended in
 10 TE, treated with RNase and reextracted with phenol-chloroform.

Analysis by fluorescence in situ hybridization (FISH). PAC or BAC clones were biotinylated by
 15 nicktranslation in the presence of biotin-14-dATP using the BioNick Labeling Kit (Gibco-BRL). FISH was performed essentially as described (Korenberg et al., Cytogenet. Cell Genet. **69**:196-200, 1995). Briefly, 400 ng of probe DNA was mixed with 8 ng of human Cot 1 DNA (Gibco-BRL)
 20 and 2 µg of sonicated salmon sperm DNA in order to suppress possible background produced from repetitive human sequences as well as yeast sequences in the probe. The probes were denatured at 75°C, preannealed at 37°C for one hour, and applied to denatured chromosome slides
 25 prepared from normal male lymphocytes (Korenberg et al., supra, 1995). Post-hybridization washes were performed at 40°C in 2X SSC/50% formamide followed by washes in 1X SSC at 50°C. Hybridized DNAs were detected with avidin-conjugated fluorescent isothiocyanate (Vector
 30 Laboratories). One amplification was performed by using biotinylated anti-avidin. For distinguishing chromosome subbands precisely, a reverse banding technique was used, which was achieved by chromomycin A3 and distamycin A double staining (Korenberg et al., supra, 1995). The
 35 color images were captured by using a Photometrics

Cooled-CCD camera and BDS image analysis software (Oncor Imaging, Inc.).

Southern blot analysis. Gel electrophoresis of DNA was carried out on 0.8% agarose gels in 1X TBE.

5 Transfer of nucleic acids to Nybond N+ nylon membrane (Amersham) was performed according to the manufacturer's instruction. Probes were labeled using RadPrime Labeling System (BRL). Hybridization was carried out at 42°C for 16 hours in 50% formamide, 5X SSPE, 5X Denhardt's 0.1% SDS, 100 mg/ml denatured salmon sperm DNA. The filters
10 were washed once in 1x SSC, 0.1% SDS at room temperature for 20 minutes, and twice in 0.1X SSC, 0.1% SDS for 20 minutes at 65°C. The blots were exposed onto X-ray film (Kodak, X-OMAT-AR).

15 *Sequencing of PAC and BAC endclones.* PAC clones were inoculated into 500 ml of LB/kanamycin and grown overnight. BAC clones were inoculated into 500 ml of LB/chloramphenicol and grown overnight. DNAs were isolated using QIAGEN columns according to the vendors
20 protocol with one additional phenol/chloroform/isoamylalcohol extraction followed by one additional chloroform/isoamylalcohol extraction. Clones were sequenced using the Gibco-BRL cycle sequencing kit with standard T7 and SP6 primers.

25 EXAMPLE 1

Construction of BAC Contig

To provide stable clones for gene isolation and sequencing initiatives in the D21S55 to MX1 region, contigs were constructed using Bacterial Artificial
30 Chromosomes (BACs) and P1 Artificial Chromosomes (PACs). BAC library construction of total human genomic DNA was performed as described (Shiyuza et al., *supra*, 1992; Kim

et al., Genomics **34**:213-218, 1996). A BAC library was screened using several YACs spanning the region; a PAC library (Iannou et al., Nature Genet. **6**:84-89, 1994) was screened using radiolabeled STS PCR products and whole
 5 BACs in gap filling initiatives.

The location of these BAC and PAC clones was confirmed by fluorescence *in situ* hybridization (FISH). Clone to clone Southern using 24 new STSs (generated from direct sequencing of BAC and PAC ends) along with 35
 10 pre-existing STSs were used to show overlaps between BACs and PACs. The STS density over the intervals covered in BACs and PACs was 1 STS every 60 kb, and 79% of the clones were positive for 2 or more STSs. Approximately
 15 3.5Mb of the 4-5Mb D21S55 to MX1 interval is covered in 85 BACs and 25 PACs representing 4-fold coverage within the contigs (Hubert et al., Genomics **41**:218-226, 1997). The minimal contig sizes as determined by counting only non-overlapping clones are: 1100 kb, 900 kb, 510 kb, 380
 20 kb and 270 kb. Insert size of BAC clones was measured by running pulse-field gel electrophoresis after digesting DNA with NotI.

EXAMPLE 2

Direct cDNA Selection

A modified direct cDNA selection technique
 25 (Yamakawa et al., Hum. Mol. Genet. **4**:709-716, 1995; Yamakawa et al., Cytogenet. Cell Genet. **74**:140-145, 1996) was applied to BAC-423A5, BAC-430F1, BAC-628H2, BAC-371H8 and PAC-31P10 (Figure 1) by using cDNA from trisomy 21 human fetal brain, and the selected fragments were then
 30 subcloned into a plasmid vector.

Total RNA was isolated from 14 week trisomy 21 fetal brain using TRI region™ (Molecular Research Center, Inc.). Poly (A)⁺ RNA was isolated using Poly (A) Quick[®] mRNA isolation kit (STRATAGENE). Double stranded cDNA
 5 was synthesized using SuperScript™ Choice System (GIBCO BRL) from 5 µg trisomy 21 fetal brain poly (A)⁺ RNA using 1 µg oligo (dT)₁₅ or 0.1 µg random hexamer. The entire synthesis reaction was purified by Gene Clean[®]II kit (BIO101, Inc.) and then kinased. Sau3AI linker was
 10 attached to the cDNA which was subsequently digested with Sau3AI. The reaction was purified using Gene Clean. MboI linker was attached to the cDNA and the reaction purified by Gene Clean (Morgan et al., supra, 1992). The synthesized product was amplified by PCR using one strand
 15 of MboI linker (5'CCTGATGCTCGAGTGAATTC3') (SEQ ID NO:4) as a primer. PCR cycling conditions were 40 cycles of 94°C/15 seconds, 60°C/23 seconds, 72°C/2 minutes in a 100 µl of 1x PCR buffer (Promega), 3 mM MgCl₂, 5.0 units of Taq polymerase (Promega), 2 µM primer and 0.2 mM dNTPs.

20 Nineteen BAC DNAs (total 2.5 µg) and 2 PAC DNAs between the region ETS2 and MX1 were prepared using QIAGEN plasmid kit and were biotinylated using Nick Translation Kit and biotin-16-dUTP (Boehringer Manneheim). 3 µg of heat denatured PCR amplified cDNA
 25 was annealed with 3 µg of heat denatured COT1 DNA (BRL) in 100µl hybridization buffer (750 mM NaCl, 50 mM NaPO₄(pH7.2), 5 mM EDTA, 5X Denhardt's, 0.05% SDS and 50% formamide) at 42°C for two hours. After prehybridization, 1.2 µg of heat denatured biotinylated
 30 BAC DNA was added and incubated at 42°C for 16 hours. cDNA-BAC DNA hybrids were precipitated with EtOH and dissolved in 60 µl of 10 mM Tris-HCl (pH 8.0), 1 mM EDTA. After addition of 40 µl 5 M NaCl, the DNA was incubated with magnetic beads (Dynabeads M-280, Dynal) at 25°C for
 35 1 hour with gentle rotating to allow attachment of the DNA to the magnetic beads. The beads were then washed

twice by pipetting in 400 μ l of 2X SSC, setting in magnet holder (MPC-E_{TM}, Dynal) for 30 seconds and removing the supernatant. Four additional washes were performed in 0.2X SSC at 68°C for 10 minutes each with transfer of the
 5 beads to new tubes at each wash. cDNAs were eluted in 100 μ l of distilled water for 10 minutes at 80°C with occasional mixing. The eluted cDNAs were amplified by PCR as described above. After twice repeating the selection procedure using magnetic beads, amplified cDNAs
 10 were digested with EcoRI and subcloned into pBlueScript KS+ (STRATAGENE). Insert DNAs were isolated from the subclones, and were analyzed by Southern hybridization and DNA sequencing.

The direct cDNA selection procedure using 19
 15 BACs and 2 PACs between ETS2 and MX1 generated a total of 145 unique cDNA fragments. Genbank and TIGR homology searches using FASTA revealed matches to ETS2, HMG14, PEP19, a Na K ATPase, Titan ESTs, MX1 region ESTs, and 14
 20 ESTs of unknown function. A cDNA library from a trisomy 21 fetal brain at 14 weeks gestation was screened using one of these unique cDNA fragments labeled "E51" (SEQ ID NO:3).

EXAMPLE 3

Isolation of human DS-CAM cDNA using cDNA Library Screening

25 A trisomy 21 human fetal brain (14 weeks of age) cDNA library was constructed using ZAP-cDNA[®] synthesis kit (STRATAGENE) which generates a unidirectional cDNA library. Briefly, double- stranded
 30 cDNA was synthesized from 5 μ g trisomy 21 fetal brain poly(A)⁺ RNA using a hybrid oligo(dT)-XhoI linker primer with 5-methyl dCTP. An EcoRI linker was attached to the cDNA which was subsequently digested with EcoRI and XhoI,

and then cloned into UNI-ZAP XR vector (STRATAGENE). The library was packaged using Gigapack[®] II Gold packaging extract. The titer of the original library was 1.1×10^6 p.f.u./package. The library was amplified once. A
 5 blue-white color assay indicated that 99% of the clones had inserts.

Screening of the trisomy 21 fetal brain cDNA library was performed using one of the 145 unique cDNA fragments labeled "E51" (SEQ ID NO:3) prepared as
 10 described above. Phages were plated to an average density of 1×10^5 per 175 cm² plate. Plaque lifts of 20 plates (2×10^6 phages) were made using duplicated nylon membranes (Hybond-N+; Amersham). Hybridized membranes were washed to final stringency of 0.2X SSC, 0.1X SDS at
 15 65°C. The filters were exposed overnight onto X-ray film.

Identification of 62 clones were made out of 2×10^6 clones in the original library. Eighteen of these positive phage clones were converted to plasmids, and
 20 their DNAs were isolated. These cDNAs were independently numbered as separate DS-CAM (Down Sndrome Cell Adhesion Molecule) clones. The length of the inserts of these clones ranged from 2.4 kb to 6.6 kb. Exon trapping (Buckler et al., Proc. Natl. Acad. Sci. USA **88**:4005-4009,
 25 1991; Church et al., Nature Genet. **6**:98-105, 1994) was also used to isolate cDNAs in the BAC and PAC contig. With this approach, three exons identified from BAC-539E7 and one from BAC-430F1 were found to identify the same sequences as those isolated by cDNA selection.

30 Sequence analysis of one of the clones, labeled DS-CAM-42, revealed a 6110 bp DNA sequence which contained a large ORF (5687 bp) as well as 3'-UTR sequence (423 bp), but the 5'UTR and start codon were not located in clone DS-CAM-42. To characterize the 5' end,

The DS-CAM1 protein contains an extracellular component at the N-terminus consisting of nine tandemly repeated Ig-like C2 type domains and a tenth Ig-like C2 domain located between domains four and five of an array of six repeated fibronectin type III domains (Figure 2). Each Ig-like C2 domain consists of approximately 100 amino acids with a pair of conserved cysteines separated by 49-56 residues. A single transmembrane domain of 22 amino acids was defined by using the TMBASE program (Hoffmann and Stoffel, Biol. Chem. Hoppe-Seyler **374**:166, 1993). The remaining 294 amino acids at the C-terminus corresponding to the cytoplasmic domain have partial homologies to the mouse M-phase inducer phosphatase 2 (Kakizuka et al., Genes Dev. **6**:578-590, 1992) in two regions, one with 34% identity and 52% similarity over 46 bp and a second with 38% identity and 52% similarity over 21 bp. The homolog of Drosophila glass gene (O'Neill et al., Proc. Natl. Acad. Sci. USA **92**:6557-6561, 1995) with

The DS-CAM1 protein contains an extracellular component at the N-terminus consisting of nine tandemly repeated Ig-like C2 type domains and a tenth Ig-like C2 domain located between domains four and five of an array of six repeated fibronectin type III domains (Figure 2). Each Ig-like C2 domain consists of approximately 100 amino acids with a pair of conserved cysteines separated by 49-56 residues. A single transmembrane domain of 22 amino acids was defined by using the TMBASE program (Hoffmann and Stoffel, Biol. Chem. Hoppe-Seyler **374**:166, 1993). The remaining 294 amino acids at the C-terminus corresponding to the cytoplasmic domain have partial homologies to the mouse M-phase inducer phosphatase 2 (Kakizuka et al., Genes Dev. **6**:578-590, 1992) in two regions, one with 34% identity and 52% similarity over 46 bp and a second with 38% identity and 52% similarity over 21 bp. The homolog of Drosophila glass gene (O'Neill et al., Proc. Natl. Acad. Sci. USA **92**:6557-6561, 1995) with

30% identity and 52% similarity over 42 bp, and the mouse delta opioid receptor (Evans et al., Science **258**:1952-1955, 1992) with 43% identity and 60% similarity over 30 bp. The putative protein contains 16 potential
 5 N-glycosylation sites.

A homology search of the predicted amino acid sequence of the 5730 bp open reading frame of DS-CAM1 (SEQ ID NO:1) to genes registered in the Genbank and the EMBL databases was conducted by using the BLAST-P program
 10 (Altschul et al., J. Mol. Biol. **215**:403-410, 1990). The predicted amino acid sequence revealed homologies to multiple proteins (Figure 4) including CAM-L1 (Moos et al., Nature **334**:701-703, 1988), BIG-1 (brain-derived immunoglobulin (Ig) superfamily molecule-1) (Yoshihara et
 15 al., Neuron **13**:415-426, 1994), DCC (deleted in colon cancer) (Fearon et al., Science **247**:49-56, 1990), and revealed DS-CAM as defining a novel class of the immunoglobulin (Ig) superfamily. Homology searches with sequences of Ig type-C2 domains and fibronectin type-III
 20 domains of the most highly related Ig-superfamily members (CAM-L1, DCC, and axonin-1) were conducted by using the FASTA program (Pearson and Lipman, Proc. Natl. Acad. Sci. USA **85**:2444-2448, 1988).

In addition, a splice variant cDNA sequence
 25 encoding a non-membrane bound isoform of DS-CAM1, referred to herein as DS-CAM2, is provided herein. Two human DS-CAM cDNA clones (DS-CAM-18 and DS-CAM-52) were found to contain identical deletions of 191 bp that occur in neighboring exons and that delete bp 5133 to 5323 of
 30 the SEQ ID NO:1 cDNA sequence encoding DS-CAM1 (Figure 3). The resulting splice variant transcript encoding DS-CAM2 (SEQ ID NO:10) is deleted for the entire transmembrane domain that is encoded by the more 3' of these exons. Further, the deletion changes the reading

frame and creates a stop codon 36 bp downstream of the deletion resulting in a soluble extracellular protein of 1571 amino acids (SEQ ID NO:11). The distal border of the resulting deletion contains the canonical AG of the RNA splicing consensus acceptor site. The proximal border contains a variant of the donor splice site consensus sequence (Jackson, Nucl. Acids Res. 19:3795-3798, 1991).

To confirm that the DS-CAM cDNA originated from the BACs and PACs in the Down syndrome region and to determine the genomic size of DS-CAM, the longest DS-CAM cDNA clones (DS-CAM-42; 6.1 kb, DS-CAM-18; 6.5 kb, DS-CAM-52; 6.6 kb) were hybridized to Southern blots containing the BAC and PAC clone contig (Figure 1). DS-CAM-42, 18 and 52 hybridized to BACs 423A5, 430F1, 628H2, 539E7, 371H8, 825E1, 593D1, 261F12, 30E4, 385B7, 388F4, and to PACs 31P10, 58D10. BACs 816F6, 116E8, 720G4, 619H8 were only positive for DS-CAM-18 and DS-CAM-52 but negative for DS-CAM-42. All other BACs shown in Figure 1 were negative. These results indicate that the DS-CAM gene spans 900 kb-1200 kb genomic DNA and covers a gap in this BAC and PAC contig indicated by an arrowhead as well as in the available YAC contigs (Korenberg et al., Genome Res. 5:427-443, 1995; Gardiner et al., Somat. Cell Mol. Genet. 21:399-414, 1995).

DS-CAM cDNA sequences were confirmed to originate from these BACs and PACs by direct sequencing of the BACs and PACs as templates using cDNA sequence-specific primers.

The map position of DS-CAM on chromosome 21q22.2-22.3 was confirmed by using clone DS-CAM-42 as a probe for fluorescence in-situ hybridization. Two independent experiments were performed and over 100 metaphase cells were evaluated. Signals were clearly seen on two chromatids of at least one chromosome in 85%

of cells. There were no other double signal sites seen in greater than 1% of cells.

EXAMPLE 4

Northern Blot Analysis Of Human DS-CAM Expression

5 Inserts containing DS-CAM cDNA were excised from the base vector by digestion with XhoI and EcoRI. After labeling using the random priming method (RadPrime Labeling System; GIBCO BRL), followed by purification using G-50 Sephadex columns (Quick Spin Column; 10 Boehringer Mannheim), the fragments were used as probes for Northern hybridization using Multiple Tissue Northern Blot (Clontech). A Northern blot assay was conducted using DS-CAM cDNA as a probe in various fetal and adult tissues including heart, brain, placenta, lung, liver, 15 skeletal muscle, kidney, and pancreas. Northern hybridization was performed by following the manufacturer's instructions. The hybridized membrane was washed at a final stringency of 0.1X SSC and 0.1X SDS at 50°C. The filter was exposed to X-ray film (Kodak X-OMAT 20 AR) at -70°C for 1-5 days.

The results of Northern analysis using human fetal tissues showed that 8.5 kb and 7.6 kb transcripts are expressed only in fetal brain and not expressed in fetal lung, fetal liver and fetal kidney. In adult 25 tissues, three transcripts of 9.7 kb, 8.5 kb, and 7.6 kb are present in the brain. Placenta shows faint bands, and the sizes are similar to those in brain. In skeletal muscle, a faint smaller band (6.5 kb) is detected. In multiple parts of the adult human brain, transcripts of 30 9.7 kb, 8.5 kb and 7.6 kb are differentially expressed. The 9.7 kb transcript is highly expressed in the substantia nigra, moderately expressed in amygdala and hippocampus, and less expressed in the whole brain. A

similar pattern is obtained using a PCR product which spans the 191 bp deletion found in clones DS-CAM-18 and DS-CAM-52 encoding the splice variant sequence corresponding to DS-CAM2. Thus, splice variant cDNA transcripts encoding a DS-CAM family of proteins are clearly contemplated by the present invention.

EXAMPLE 5

RT-PCR Assays Of Human DS-CAM Expression

Reverse-transcriptase polymerase chain reaction (RT-PCR) assays verses cDNA libraries of various human tissues were conducted using primers numbered B9-131F (SEQ ID NO:5) and B9-131R (SEQ ID NO:6). The results demonstrated expression of human DS-CAM mRNA in fetal and adult brain, and fetal kidney. In addition, a breast carcinoma cell line showed expression of human DS-CAM mRNA.

The cDNAs from 13 independent human fetal and adult sources were analyzed by PCR using primer pairs that flanked the alternatively spliced region that results in a 191 base pair deletion of nucleotides 5133-5323 of the DS-CAM1 cDNA set forth in SEQ ID NO:1. The primers were designed to generate products of different sizes for each of the two alternatively spliced transcripts: 536 bp corresponding to the non-deleted DS-CAM-1 transcript and 345 bp corresponding to the deleted DS-CAM2 transcripts. The analyses included adult samples from amygdala (24 years), skeletal muscle (36 years) and three independent lymphoblastoid cell lines. Fetal samples included whole brain of a trisomy 21 fetus (14 weeks), four from whole brain (4.5-13 weeks), one from temporal lobe (28 weeks) and two from heart (4.5 and 13 weeks). The results indicate that all fetal and adult samples produced two bands corresponding to PCR products

of the predicted sizes which indicates the expression of two alternatively spliced transcripts.

EXAMPLE 6

Isolation of mouse DS-CAM cDNA clones

5 A mouse brain cDNA library was prepared from 19 week old female C57 Black/6 mice in the Uni-ZAP XR Vector (STRATAGENE). The cDNAs were oligo-dT primed and cloned unidirectionally into the EcoRI and XhoI sites of the vector. The average insert size is 1.0 kb. The library
10 was screened using a human DS-CAM cDNA clone as a probe. Two partial mouse DS-CAM cDNA clones were isolated and sequenced. The combined nucleotide sequences of these clones are set forth in SEQ ID NO:7, SEQ ID NO:8 and SEQ ID NO:9, and were found to represent the 5', middle
15 and 3' portions, respectively, of cDNA encoding a mouse DS-CAM.

EXAMPLE 7

Hybridization analysis of DS-CAM cDNA in mouse tissues

BALB/c and C57BL/6 x DBA/2 embryos, fetuses and
20 postnatal brains were fixed and embedded as described in detail in Lyons et al., (J. Neurosci. **15**:5727-5738, 1995). Embryos were fixed in 4% paraformaldehyde in phosphate buffered saline (PBS) overnight, dehydrated and infiltrated with paraffin. Five to seven micron serial
25 sections were mounted on gelatinized slides. Two sections were mounted/slide, deparaffinized in xylene, rehydrated and post-fixed. The sections were digested with proteinase K, post-fixed, treated with tri-ethanolamine/acetic anhydride, washed and dehydrated.
30 cRNA probes were prepared from DS-CAM-M-14. The plasmid was linearized with XbaI and T7 polymerase was used to

generate the antisense cRNA. The plasmid was linearized with KpnI and T3 polymerase was used to generate the sense control cRNA. The cRNA transcripts were synthesized according to manufacturer's conditions (STRATAGENE) and labeled with ^{35}S -UTP (>1000 Ci/mmol; Amersham). cRNA transcripts larger than 100 nucleotides were subjected to alkali hydrolysis to give a mean size of 70 bases for efficient hybridization.

Sections were hybridized overnight at 52°C in 50% deionized formamide, 0.3M NaCl, 20 mM Tris-HCl pH 7.4, 5 mM EDTA, 10 mM NaPO_4 , 10% dextran sulfate, 1x Denhardt's, 50 $\mu\text{g}/\text{ml}$ total yeast RNA, and 50-75,000 cpm/ μl ^{35}S -labeled cRNA probe. The tissue was subjected to stringent washing at 65°C in 50% formamide, 2X SSC, 10 mM DTT and washed in PBS before treatment with 20 $\mu\text{g}/\text{ml}$ RNase A at 37°C for 30 minutes. Following washes in 2X SSC and 0.1X SSC for 10 minutes at 37°C , the slides were dehydrated and dipped in Kodak NTB-2 nuclear track emulsion and exposed for 2-3 weeks in light-tight boxes with desiccant at 4°C . Photographic development was carried out in Kodak D-19. Slides were counterstained lightly with toluidine blue and analyzed using both light- and darkfield optics of a Zeiss Axiophot microscope. Sense control cRNA probes (identical to the mRNAs) always gave background levels of hybridization signal. Embryonic structures were identified with the help of the following atlases: Rugh (The Mouse: Its Reproduction and Development. Oxford Univ. Press, Oxford, UK, 1990), Kaufman (The Atlas of Mouse Development. Acad. Press, New York, NY, 1992), and Altman and Bayer (supra, 1995).

Tissue in situ hybridization analysis was performed using a mouse cDNA as a probe on sections of normal mouse embryos from days 8.5-17.5 post coitum (pc) as well as in newborn, two weeks and adult brains as

5 branchial arches, suggestive of migrating neural crest cells. At 10.5 days pc, the trigeminal ganglia (neural crest derived) begin to express the transcript and expression within the branchial arches was more evident.

10 throughout the brain. The transcript was found within
the regions of the nervous system that differentiate
earliest during development (Altman and Bayer, supra,
1995). In the brain, this includes the ventral-most
regions, such as the thalamus and medulla. Some
15 expression was detected within the olfactory epithelium.
Expression within the neural tube begins in two areas:
the ventrolateral (corresponding to the areas in which
the motor neurons differentiate) and the lateral gray
columns (that later form commissural neurons) (Leber et
20 al., J Neurosci. **15**:1236-1248, 1990). The dorsal root
ganglia (neural crest derived) expressed the transcript
at 11.5 days pc. The trigeminal ganglia show higher
levels at 11.5 days pc than they did at 10.5 days.
Migrating neural crest can be seen within the maxilla,
25 the mandibular arch, and in the developing gut. Signal
was observed within the mesenchyme surrounding the
umbilical vein and artery.

30 exhibits expression of the transcript, including a larger portion of midbrain, the pontine areas, the basal ganglia and the outermost layer of cortex. Neurons in this layer have undergone mitosis in the subependymal layer of the cortex and migrated into the mantle layer of the cerebral

cortex as differentiated cells (Smart et al., J. Comp. Neurol. **116**:325-347, 1961).

At 13.5 days pc, expression was seen throughout most of the brain. The outermost layer of the gut also
5 appears to be expressing at this stage; these cells are neural crest derived and form the myenteric ganglia. At 15.5 and 16.5 days pc, most of the neural crest derived neural structures have some expression. For example, the
10 structures at the base of the vibrissae, the pancreatic ganglia, the heart ganglion, the enteric nervous system, and the sympathetic trunk all express the transcript.

There is no expression within the umbilicus at this stage. Two non-neuronal structures express this
15 gene, the gonad and the annulus fibrosus of the intervertebral disk. The olfactory bulb exhibits signal both in the granule cells and within the tufted mitral cells. Within the newborn brain, the transcript was expressed most extensively within the differentiating
20 regions such as the septal area, olfactory bulb, inferior colliculus and hippocampus. In the adult brain, the gene was expressed in many areas including amygdala, cortex, hippocampus and thalamus. In the adult cerebellum the transcripts were detected in the Purkinje cell layer and
25 in the deep cerebellar nuclei.

While the invention has been described in detail with reference to certain preferred embodiments thereof, it will be understood that modifications and variations are within the spirit and scope of that which
30 is described and claimed.

Summary of Sequences

SEQ ID NO:1 is the nucleic acid sequence (and the deduced amino acid sequence) of cDNA encoding a novel human DS-CAM1 protein of the present invention.

- 5 SEQ ID NO:2 is the deduced amino acid sequence of a human DS-CAM1 protein of the present invention.

SEQ ID NO:3 is the cDNA probe (labeled "E51") used to isolate cDNA encoding human DS-CAM.

SEQ ID NO:4 is an MboI linker sequence.

- 10 SEQ ID NO:5 is a primer labeled B9-131F used in the RT-PCR assay described in Example 5.

SEQ ID NO:6 is a primer labeled B9-131R used in the RT-PCR assay described in Example 5.

- 15 SEQ ID NO:7 is the 5' region of a partial mouse-derived cDNA clone encoding an invention DS-CAM protein.

SEQ ID NO:8 is the middle region of a partial mouse-derived cDNA clone encoding an invention DS-CAM protein.

- 20 SEQ ID NO:9 is the 3' region of a partial mouse-derived cDNA clone encoding an invention DS-CAM protein.

SEQ ID NO:10 is the nucleic acid sequence (and the deduced amino acid sequence) of cDNA encoding a novel human DS-CAM2 protein of the present invention.

- 25 SEQ ID NO:11 is the deduced amino acid sequence of a human DS-CAM2 protein of the present invention, which is a splice variant of DS-CAM1 (SEQ ID NO:2).

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (I) APPLICANT: Korenberg, Julie R.
- (ii) TITLE OF INVENTION: NUCLEIC ACID ENCODING DS-CAM
PROTEINS AND PRODUCTS RELATED THERETO
- (iii) NUMBER OF SEQUENCES: 11
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Campbell and Flores
 - (B) STREET: 4370 La Jolla Village Drive, Suite 700
 - (C) CITY: San Diego
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 92122
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 60/029,322
 - (B) FILING DATE: 25-OCT-1996
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Ramos, Robert T.
 - (B) REGISTRATION NUMBER: 37,915
 - (C) REFERENCE/DOCKET NUMBER: P-CE 2817
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 619-535-9001
 - (B) TELEFAX: 619-535-8949

(2) INFORMATION FOR SEQ ID NO:1:

- (I) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 6604 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: both
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 453..6185

"CE" of "16695580"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

TGACTGAGGC CGGAGCACGG CAAAGATGAG CCTGCCCGCC CGCCTGCTGC CTGGATGCGG	60
AGGGTGAGGG CTGGCGCACG GGAGGCCGCT GGCTGCGCAT TCTGGGCGCC GAGTGCCCGG	120
GATGAGCTCA CGCCCGCGTC TGC GGCTCTC TCCACCTGCC GACCTGCCGG GGGCCCACTG	180
AGCTGACGGC GCACCTGGGC TCCGGCCGCA GCGTGGGGCG CGGCGCCCGG GAGCAGGTGT	240
GCAGGAGCGC AGCGCGCGGC GAGCGCAGCC CTCGCTCCGG AGCCCGGCCG CGCCGCGTGC	300
CCGGGCGGGCT AGGCAGCGGC GGC GGCGGCG GCGGGCGGCG GGCGGGCGGC GGCCCCCGGG	360
CAGGTGCCGA GCGGCGAGCG GAGCCGGGCC GGGCGGAGCG CGGGGGGCGA GGCCGGCGCG	420
TCGCTCGCGG GAGGCCGGGG AGCGGCAGGG GC ATG TGG ATA CTG GCT CTC TCC	473
Met Trp Ile Leu Ala Leu Ser	
1 5	
TTG TTC CAG AGC TTC GCG AAT GTT TTC AGT GAA GAC CTA CAC TCC AGC	521
Leu Phe Gln Ser Phe Ala Asn Val Phe Ser Glu Asp Leu His Ser Ser	
10 15 20	
CTC TAC TTT GTC AAT GCA TCT CTG CAA GAG GTA GTG TTT GCC AGC ACC	569
Leu Tyr Phe Val Asn Ala Ser Leu Gln Glu Val Val Phe Ala Ser Thr	
25 30 35	
ACG GGG ACT CTG GTG CCC TGC CCC GCA GCA GGC ATC CCT CCT GTG ACT	617
Thr Gly Thr Leu Val Pro Cys Pro Ala Ala Gly Ile Pro Pro Val Thr	
40 45 50 55	
CTC AGA TGG TAC CTA GCC ACG GGC GAG GAG ATC TAC GAT GTC CCC GGG	665
Leu Arg Trp Tyr Leu Ala Thr Gly Glu Glu Ile Tyr Asp Val Pro Gly	
60 65 70	
ATC CGC CAC GTC CAC CCC AAC GGC ACT CTC CAA ATT TTC CCC TTC CCT	713
Ile Arg His Val His Pro Asn Gly Thr Leu Gln Ile Phe Pro Phe Pro	
75 80 85	
CCT TCA AGC TTC AGT ACC TTA ATC CAT GAT AAT ACT TAT TAT TGC ACA	761
Pro Ser Ser Phe Ser Thr Leu Ile His Asp Asn Thr Tyr Tyr Cys Thr	
90 95 100	
GCT GAA AAT CCT TCA GGG AAA ATT AGA AGT CAG GAT GTC CAC ATC AAG	809
Ala Glu Asn Pro Ser Gly Lys Ile Arg Ser Gln Asp Val His Ile Lys	
105 110 115	
GCT GTT TTA CGG GAG CCC TAT ACA GTC CGT GTG GAG GAC CAG AAA ACC	857
Ala Val Leu Arg Glu Pro Tyr Thr Val Arg Val Glu Asp Gln Lys Thr	
120 125 130 135	
ATG AGA GGC AAT GTT GCG GTC TTC AAG TGC ATT ATC CCC TCC TCG GTG	905
Met Arg Gly Asn Val Ala Val Phe Lys Cys Ile Ile Pro Ser Ser Val	
140 145 150	
GAG GCG TAC ATC ACT GTC GTC TCA TGG GAG AAA GAC ACT GTT TCA CTT	953
Glu Ala Tyr Ile Thr Val Val Ser Trp Glu Lys Asp Thr Val Ser Leu	
155 160 165	
GTC TCA GGA TCT AGA TTT CTC ATC ACA TCC ACG GGA GCC TTG TAT ATT	1001
Val Ser Gly Ser Arg Phe Leu Ile Thr Ser Thr Gly Ala Leu Tyr Ile	
170 175 180	

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1865	AGC Ser	CAG Gln	ATG Met	ATC Ile 460	ACG Thr	TCG Ser	GAG Glu	GGG Gly	AAC Asn	GTG Val 465	GTC Val	AGC Ser	TAC Tyr	CTG Leu	AAC Asn 470	ATC Ile
1913	TCC Ser	AGC Ser	TCC Ser	CAG Gln 475	GTC Val	CGG Arg	GAC Asp	GGG Gly	GGA Gly 480	GTC Val	TAC Tyr	CGC Arg	TGC Cys	ACT Thr 485	GCC Ala	AAC Asn
1961	AAC Asn	TCG Ser	GCG Ala 490	GGA Gly	GTC Val	GTC Val	CTG Leu	TAC Tyr 495	CAG Gln	GCT Ala	CGA Arg	ATA Ile	AAC Asn 500	GTA Val	AGA Arg	GGG Gly
2009	CCT Pro	GCA Ala 505	AGC Ser	ATT Ile	CGA Arg	CCA Pro	ATG Met 510	AAA Lys	AAC Asn	ATC Ile	ACA Thr	GCA Ala 515	ATA Ile	GCA Ala	GGA Gly	CGG Arg
2057	GAC Asp 520	ACA Thr	TAC Tyr	ATT Ile	CAC His	TGT Cys 525	CGT Arg	GTG Val	ATT Ile	GGC Gly	TAT Tyr 530	CCG Pro	TAT Tyr	TAC Tyr	TCC Ser	ATT Ile 535
2105	AAA Lys	TGG Trp	TAC Tyr	AAG Lys	AAC Asn 540	TCT Ser	AAC Asn	CTG Leu	CTT Leu	CCT Pro 545	TTC Phe	AAC Asn	CAC His	CGC Arg	CAA Gln 550	GTG Val
2153	GCA Ala	TTT Phe	GAG Glu	AAC Asn 555	AAT Asn	GGA Gly	ACT Thr	CTT Leu	AAA Lys 560	CTT Leu	TCA Ser	GAT Asp	GTG Val	CAA Gln 565	AAG Lys	GAA Glu
2201	GTG Val	GAC Asp	GAG Glu	GGG Gly 570	GAG Glu	TAC Tyr	ACG Thr	TGC Cys 575	AAC Asn	GTG Val	TTG Leu	GTT Val	CAA Gln 580	CCA Pro	CAA Gln	CTC Leu
2249	TCC Ser	ACC Thr 585	AGC Ser	CAG Gln	AGC Ser	GTC Val	CAC His 590	GTG Val	ACC Thr	GTG Val	AAA Lys 595	GTT Val	CCG Pro	CCT Pro	TTC Phe	ATA Ile
2297	CAA Gln 600	CCC Pro	TTT Phe	GAG Glu	TTT Phe	CCA Pro 605	AGA Arg	TTC Phe	TCC Ser	ATT Ile	GGG Gly 610	CAG Gln	CGG Arg	GTC Val	TTC Phe	ATC Ile 615
2345	CCC Pro	TGT Cys	GTT Val	GTG Val 620	GTC Val	TCA Ser	GGG Gly	GAC Asp	TTA Leu	CCC Pro 625	ATC Ile	ACG Thr	ATC Ile	ACC Thr	TGG Trp 630	CAG Gln
2393	AAG Lys	GAT Asp	GGC Gly	CGG Arg 635	CCA Pro	ATC Ile	CCT Pro	GGG Gly	AGC Ser 640	CTT Leu	GGG Gly	GTG Val	ACC Thr	ATT Ile 645	GAC Asp	AAT Asn
2441	ATT Ile	GAC Asp	TTC Phe 650	ACG Thr	AGC Ser	TCC Ser	TTG Leu	AGG Arg 655	ATT Ile	TCC Ser	AAT Asn	CTC Leu	TCG Ser	CTC Leu	ATG Met	CAC His
2489	AAT Asn	GGG Gly 665	AAT Asn	TAC Tyr	ACC Thr	TGC Cys	ATA Ile 670	GCC Ala	CGG Arg	AAT Asn	GAG Glu	GCC Ala 675	GCC Ala	GCT Ala	GTG Val	GAG Glu
2537	CAC His 680	CAA Gln	AGC Ser	CAG Gln	TTG Leu	ATT Ile 685	GTC Val	AGA Arg	GTT Val	CCT Pro	CCC Pro 690	AAG Lys	TTT Phe	GTG Val	GTT Val	CAG Gln 695

CCA	CGG	GAC	CAG	GAC	GGG	ATT	TAT	GGC	AAA	GCA	GTC	ATC	CTC	AAT	TGT	2585
Pro	Arg	Asp	Gln	Asp	Gly	Ile	Tyr	Gly	Lys	Ala	Val	Ile	Leu	Asn	Cys	
			700						705					710		
TCT	GCT	GAG	GGT	TAC	CCT	GTA	CCT	ACC	ATC	GTG	TGG	AAA	TTC	TCT	AAA	2633
Ser	Ala	Glu	Gly	Tyr	Pro	Val	Pro	Thr	Ile	Val	Trp	Lys	Phe	Ser	Lys	
			715					720					725			
GGT	GCT	GGG	GTT	CCC	CAG	TTC	CAG	CCA	ATT	GCC	CTA	AAT	GGC	CGA	ATC	2681
Gly	Ala	Gly	Val	Pro	Gln	Phe	Gln	Pro	Ile	Ala	Leu	Asn	Gly	Arg	Ile	
		730					735					740				
CAA	GTT	CTC	AGC	AAT	GGG	TCG	TTG	CTG	ATC	AAG	CAT	GTC	GTG	GAG	GAA	2729
Gln	Val	Leu	Ser	Asn	Gly	Ser	Leu	Leu	Ile	Lys	His	Val	Val	Glu	Glu	
	745					750					755					
GAC	AGT	GGC	TAC	TAC	CTC	TGC	AAG	GTC	AGC	AAC	GAT	GTG	GGC	GCA	GAC	2777
Asp	Ser	Gly	Tyr	Tyr	Leu	Cys	Lys	Val	Ser	Asn	Asp	Val	Gly	Ala	Asp	
760					765					770					775	
GTC	AGC	AAG	TCC	ATG	TAC	CTC	ACG	GTT	AAA	ATT	CCT	GCG	ATG	ATA	ACA	2825
Val	Ser	Lys	Ser	Met	Tyr	Leu	Thr	Val	Lys	Ile	Pro	Ala	Met	Ile	Thr	
				780					785					790		
TCC	TAT	CCA	AAT	ACT	ACC	CTG	GCC	ACG	CAG	GGG	CAG	AAA	AAG	GAG	ATG	2873
Ser	Tyr	Pro	Asn	Thr	Thr	Leu	Ala	Thr	Gln	Gly	Gln	Lys	Lys	Glu	Met	
			795					800					805			
AGC	TGC	ACG	GCG	CAT	GGT	GAG	AAG	CCC	ATT	ATA	GTC	CGC	TGG	GAG	AAG	2921
Ser	Cys	Thr	Ala	His	Gly	Glu	Lys	Pro	Ile	Ile	Val	Arg	Trp	Glu	Lys	
		810					815					820				
GAG	GAC	CGA	ATC	ATT	AAC	CCT	GAG	ATG	GCC	CGT	TAT	CTT	GTG	TCC	ACC	2969
Glu	Asp	Arg	Ile	Ile	Asn	Pro	Glu	Met	Ala	Arg	Tyr	Leu	Val	Ser	Thr	
	825					830					835					
AAG	GAG	GTG	GGA	GAA	GAG	GTG	ATT	TCT	ACT	CTG	CAG	ATT	TTG	CCA	ACT	3017
Lys	Glu	Val	Gly	Glu	Glu	Val	Ile	Ser	Thr	Leu	Gln	Ile	Leu	Pro	Thr	
840					845					850					855	
GTG	AGA	GAA	GAT	TCT	GGT	TTC	TTT	TCC	TGC	CAT	GCT	ATT	AAT	TCT	TAT	3065
Val	Arg	Glu	Asp	Ser	Gly	Phe	Phe	Ser	Cys	His	Ala	Ile	Asn	Ser	Tyr	
				860					865					870		
GGG	GAG	GAC	CGT	GGA	ATA	ATT	CAG	CTC	ACA	GTG	CAA	GAG	CCC	CCA	GAC	3113
Gly	Glu	Asp	Arg	Gly	Ile	Ile	Gln	Leu	Thr	Val	Gln	Glu	Pro	Pro	Asp	
			875					880					885			
CCT	CCC	GAA	ATT	GAG	ATC	AAA	GAT	GTC	AAA	GCA	CGC	ACA	ATT	ACG	CTC	3161
Pro	Pro	Glu	Ile	Glu	Ile	Lys	Asp	Val	Lys	Ala	Arg	Thr	Ile	Thr	Leu	
		890					895					900				
AGG	TGG	ACC	ATG	GGG	TTT	GAT	GGA	AAC	AGT	CCC	ATC	ACA	GGC	TAC	GAT	3209
Arg	Trp	Thr	Met	Gly	Phe	Asp	Gly	Asn	Ser	Pro	Ile	Thr	Gly	Tyr	Asp	
	905					910					915					
ATT	GAA	TGC	AAA	AAT	AAA	TCA	GAC	TCC	TGG	GAT	TCT	GCT	CAG	AGA	ACC	3257
Ile	Glu	Cys	Lys	Asn	Lys	Ser	Asp	Ser	Trp	Asp	Ser	Ala	Gln	Arg	Thr	
920					925					930					935	
AAA	GAT	GTT	TCC	CCT	CAG	CTG	AAC	TCG	GCC	ACC	ATC	ATT	GAT	ATC	CAC	3305
Lys	Asp	Val	Ser	Pro	Gln	Leu	Asn	Ser	Ala	Thr	Ile	Ile	Asp	Ile	His	
				940					945					950		

CCT TCC TCC ACC TAC AGC ATC CGC ATG TAC GCC AAG AAC CGG ATT GGC Pro Ser Ser Thr Tyr Ser Ile Arg Met Tyr Ala Lys Asn Arg Ile Gly 955 960 965	3353
AAG AGC GAG CCC AGC AAC GAG CTC ACC ATC ACG GCG GAC GAG GCA GCT Lys Ser Glu Pro Ser Asn Glu Leu Thr Ile Thr Ala Asp Glu Ala Ala 970 975 980	3401
CCT GAT GGT CCA CCT CAG GAA GTT CAC CTG GAG CCT ATA TCA TCT CAG Pro Asp Gly Pro Pro Gln Glu Val His Leu Glu Pro Ile Ser Ser Gln 985 990 995	3449
AGC ATC AGG GTC ACA TGG AAG GCT CCC AAG AAA CAT TTG CAA AAT GGG Ser Ile Arg Val Thr Trp Lys Ala Pro Lys Lys His Leu Gln Asn Gly 1000 1005 1010 1015	3497
ATT ATC CGT GGC TAC CAA ATA GGT TAC CGA GAG TAC AGC ACT GGG GGT Ile Ile Arg Gly Tyr Gln Ile Gly Tyr Arg Glu Tyr Ser Thr Gly Gly 1020 1025 1030	3545
AAC TTC CAA TTC AAC ATT ATC AGT GTC GAC ACC AGC GGG GAC AGT GAG Asn Phe Gln Phe Asn Ile Ile Ser Val Asp Thr Ser Gly Asp Ser Glu 1035 1040 1045	3593
GTT TAC ACC CTG GAC AAC CTG AAT AAG TTC ACT CAG TAC GGC CTG GTG Val Tyr Thr Leu Asp Asn Leu Asn Lys Phe Thr Gln Tyr Gly Leu Val 1050 1055 1060	3641
GTG CAG GCC TGT AAC CGG GCC GGC ACG GGG CCT TCT TCT CAG GAA ATC Val Gln Ala Cys Asn Arg Ala Gly Thr Gly Pro Ser Ser Gln Glu Ile 1065 1070 1075	3689
ATC ACC ACC ACT CTC GAG GAT GTG CCC AGT TAC CCC CCC GAA AAT GTC Ile Thr Thr Thr Leu Glu Asp Val Pro Ser Tyr Pro Pro Glu Asn Val 1080 1085 1090 1095	3737
CAA GCC ATA GCA ACA TCA CCA GAA AGC ATA TCA ATA TCC TGG TCC ACA Gln Ala Ile Ala Thr Ser Pro Glu Ser Ile Ser Ile Ser Trp Ser Thr 1100 1105 1110	3785
CTT TCC AAG GAA GCC TTG AAT GGA ATT CTC CAG GGG TTC AGA GTC ATT Leu Ser Lys Glu Ala Leu Asn Gly Ile Leu Gln Gly Phe Arg Val Ile 1115 1120 1125	3833
TAC TGG GCC AAC CTC ATG GAC GGA GAG CTG GGT GAG ATT AAA AAC ATC Tyr Trp Ala Asn Leu Met Asp Gly Glu Leu Gly Glu Ile Lys Asn Ile 1130 1135 1140	3881
ACC ACC ACA CAG CCT TCA CTG GAG CTG GAC GGG CTG GAA AAG TAC ACC Thr Thr Thr Gln Pro Ser Leu Glu Leu Asp Gly Leu Glu Lys Tyr Thr 1145 1150 1155	3929
AAC TAC AGC ATC CAG GTG CTG GCC TTC ACC CGC GCA GGA GAC GGG GTC Asn Tyr Ser Ile Gln Val Leu Ala Phe Thr Arg Ala Gly Asp Gly Val 1160 1165 1170 1175	3977
AGG AGT GAG CAG ATC TTC ACC CGG ACC AAA GAG GAT GTT CCA GGT CCT Arg Ser Glu Gln Ile Phe Thr Arg Thr Lys Glu Asp Val Pro Gly Pro 1180 1185 1190	4025
CCC GCG GGT GTG AAG GCA GCG GCG GCC TCA GCC TCC ATG GTC TTT GTG Pro Ala Gly Val Lys Ala Ala Ala Ala Ser Ala Ser Met Val Phe Val 1195 1200 1205	4073

TCC Ser	TGG Trp	CTT Leu	CCC Pro	CCT Pro	CTC Leu	AAG Lys	CTG Leu	AAC Asn	GGC Gly	ATC Ile	ATC Ile	CGA Arg	AAG Lys	TAC Tyr	ACT Thr	4121
1210																
GTA Val	TTC Phe	TGC Cys	TCC Ser	CAC His	CCC Pro	TAT Tyr	CCC Pro	ACA Thr	GTG Val	ATC Ile	AGC Ser	GAG Glu	TTT Phe	GAG Glu	GCC Ala	4169
1225																
1230																
1235																
TCT Ser	CCC Pro	GAC Asp	TCG Ser	TTT Phe	TCC Ser	TAC Tyr	AGA Arg	ATT Ile	CCC Pro	AAC Asn	CTG Leu	AGT Ser	AGG Arg	AAT Asn	CGT Arg	4217
1240																
1245																
1250																
1255																
CAG Gln	TAC Tyr	AGC Ser	GTC Val	TGG Trp	GTG Val	GTG Val	GCT Ala	GTT Val	ACT Thr	TCA Ser	GCC Ala	GGA Gly	AGA Arg	GGC Gly	AAC Asn	4265
1260																
1265																
1270																
AGC Ser	AGT Ser	GAA Glu	ATC Ile	ATC Ile	ACA Thr	GTC Val	GAG Glu	CCA Pro	CTA Leu	GCA Ala	AAA Lys	GCT Ala	CCT Pro	GCA Ala	CGA Arg	4313
1275																
1280																
1285																
ATC Ile	CTG Leu	ACC Thr	TTC Phe	AGT Ser	GGG Gly	ACA Thr	GTG Val	ACT Thr	ACT Thr	CCA Pro	TGG Trp	ATG Met	AAA Lys	GAC Asp	ATT Ile	4361
1290																
1295																
1300																
GTC Val	TTG Leu	CCT Pro	TGT Cys	AAG Lys	GCT Ala	GTT Val	GGG Gly	GAC Asp	CCT Pro	TCT Ser	CCT Pro	GCA Ala	GTC Val	AAA Lys	TGG Trp	4409
1305																
1310																
1315																
ATG Met	AAA Lys	GAC Asp	AGT Ser	AAC Asn	GGG Gly	ACA Thr	CCC Pro	AGT Ser	CTA Leu	GTA Val	ACG Thr	ATT Ile	GAT Asp	GGG Gly	CGG Arg	4457
1320																
1325																
1330																
1335																
AGG Arg	AGC Ser	ATC Ile	TTT Phe	AGC Ser	AAC Asn	GGA Gly	AGC Ser	TTC Phe	ATT Ile	ATT Ile	CGC Arg	ACG Thr	GTG Val	AAA Lys	GCA Ala	4505
1340																
1345																
1350																
GAA Glu	GAC Asp	TCC Ser	GGC Gly	TAT Tyr	TAC Tyr	AGC Ser	TGC Cys	ATT Ile	GCC Ala	AAT Asn	AAC Asn	AAC Asn	TGG Trp	GGG Gly	TCT Ser	4553
1355																
1360																
1365																
GAT Asp	GAA Glu	ATT Ile	ATT Ile	TTA Leu	AAC Asn	TTA Leu	CAA Gln	GTA Val	CAA Gln	GTT Val	CCA Pro	CCA Pro	GAT Asp	CAG Gln	CCT Pro	4601
1370																
1375																
1380																
CGG Arg	CTT Leu	ACA Thr	GTC Val	TCC Ser	AAG Lys	ACC Thr	ACG Thr	TCT Ser	TCC Ser	TCC Ser	ATC Ile	ACC Thr	CTT Leu	TCT Ser	TGG Trp	4649
1385																
1390																
1395																
CTC Leu	CCT Pro	GGA Gly	GAC Asp	AAC Asn	GGG Gly	GGC Gly	AGC Ser	TCT Ser	ATC Ile	AGA Arg	GGA Gly	TAC Tyr	ATA Ile	CTG Leu	CAG Gln	4697
1400																
1405																
1410																
1415																
TAC Tyr	TCC Ser	GAG Glu	GAC Asp	AAT Asn	AGT Ser	GAG Glu	CAG Gln	TGG Trp	GGG Gly	AGT Ser	TTT Phe	CCA Pro	ATC Ile	AGC Ser	CCC Pro	4745
1420																
1425																
1430																
AGC Ser	GAA Glu	CGT Arg	TCC Ser	TAT Tyr	CGC Arg	TTG Leu	GAA Glu	AAT Asn	CTC Leu	AAA Lys	TGT Cys	GGG Gly	ACT Thr	TGG Trp	TAT Tyr	4793
1435																
1440																
1445																
AAG Lys	TTC Phe	ACA Thr	CTG Leu	ACA Thr	GCC Ala	CAA Gln	AAT Asn	GGA Gly	GTG Val	GGC Gly	CCA Pro	GGG Gly	CGC Arg	ATA Ile	AGT Ser	4841
1450																
1455																
1460																

GAA Glu	ATC Ile	ATA Ile	GAA Glu	GCA Ala	AAG Lys	ACC Thr	TTA Leu	GGA Gly	AAA Lys	GAG Glu	CCC Pro	CAG Gln	TTC Phe	TCA Ser	AAG Lys	4889
1465			1470			1475										
GAG Glu	CAG Gln	GAG Glu	CTG Leu	TTT Phe	GCC Ala	AGC Ser	ATC Ile	AAC Asn	ACC Thr	ACA Thr	CGC Arg	GTG Val	AGG Arg	CTG Leu	AAC Asn	4937
1480			1485			1490			1495							
CTC Leu	ATT Ile	GGC Gly	TGG Trp	AAT Asn	GAT Asp	GGC Gly	GGC Gly	TGC Cys	CCC Pro	ATC Ile	ACC Thr	TCC Ser	TTC Phe	ACA Thr	CTA Leu	4985
1500				1505				1510								
GAG Glu	TAC Tyr	AGG Arg	CCC Pro	TTT Phe	GGG Gly	ACC Thr	ACA Thr	GTT Val	TGG Trp	ACC Thr	ACA Thr	GCT Ala	CAG Gln	AGG Arg	ACC Thr	5033
1515			1520			1525										
TCT Ser	CTC Leu	TCC Ser	AAG Lys	TCC Ser	TAC Tyr	ATC Ile	CTG Leu	TAT Tyr	GAC Asp	CTG Leu	CAG Gln	GAA Glu	GCC Ala	ACC Thr	TGG Trp	5081
1530			1535			1540										
TAT Tyr	GAG Glu	CTG Leu	CAG Gln	ATG Met	CGG Arg	GTG Val	TGC Cys	AAC Asn	AGT Ser	GCG Ala	GGC Gly	TGC Cys	GCG Ala	GAG Glu	AAG Lys	5129
1545			1550			1555										
CAG Gln	GCC Ala	AAC Asn	TTC Phe	GCT Ala	ACG Thr	CTG Leu	AAC Asn	TAC Tyr	GAT Asp	GGC Gly	AGT Ser	ACA Thr	ATT Ile	CCT Pro	CCA Pro	5177
1560			1565			1570			1575							
CTC Leu	ATT Ile	AAG Lys	TCA Ser	GTT Val	GTC Val	CAA Gln	AAC Asn	GAA Glu	GAA Glu	GGG Gly	CTG Leu	ACG Thr	ACC Thr	AAC Asn	GAG Glu	5225
1580			1585			1590										
GGG Gly	CTC Leu	AAG Lys	ATG Met	CTG Leu	GTG Val	ACC Thr	ATC Ile	TCC Ser	TGT Cys	ATC Ile	CTG Leu	GTG Val	GGG Gly	GTC Val	TTG Leu	5273
1595			1600			1605										
CTG Leu	CTG Leu	TTT Phe	GTG Val	CTC Leu	CTG Leu	CTG Leu	GTT Val	GTG Val	CGG Arg	AGG Arg	AGG Arg	CGG Arg	CGG Arg	GAG Glu	CAG Gln	5321
1610			1615			1620										
AGG Arg	CTA Leu	AAG Lys	AGG Arg	CTG Leu	CGA Arg	GAT Asp	GCA Ala	AAG Lys	AGT Ser	TTA Leu	GCT Ala	GAA Glu	ATG Met	CTC Leu	ATG Met	5369
1625			1630			1635										
AGT Ser	AAG Lys	AAT Asn	ACC Thr	CGG Arg	ACT Thr	TCA Ser	GAT Asp	ACG Thr	TTA Leu	AGC Ser	AAG Lys	CAA Gln	CAG Gln	CAG Gln	ACC Thr	5417
1640			1645			1650			1655							
CTG Leu	CGA Arg	ATG Met	CAC His	ATC Ile	GAC Asp	ATA Ile	CCC Pro	AGG Arg	GCT Ala	CAG Gln	CTT Leu	TTG Leu	ATT Ile	GAA Glu	GAG Glu	5465
1660			1665			1670										
AGA Arg	GAC Asp	ACG Thr	ATG Met	GAG Glu	ACC Thr	ATT Ile	GAT Asp	GAT Asp	CGC Arg	TCC Ser	ACG Thr	GTT Val	CTG Leu	TTG Leu	ACG Thr	5513
1675			1680			1685										
GAT Asp	GCT Ala	GAC Asp	TTT Phe	GGA Gly	GAG Glu	GCA Ala	GCT Ala	AAG Lys	CAG Gln	AAG Lys	TCC Ser	CTG Leu	ACG Thr	GTC Val	ACT Thr	5561
1690			1695			1700										
CAC His	ACG Thr	GTC Val	CAT His	TAC Tyr	CAA Gln	TCG Ser	GTG Val	TCT Ser	CAG Gln	GCC Ala	ACT Thr	GGG Gly	CCC Pro	TTA Leu	GTG Val	5609
1705			1710			1715										

GAT GTT TCA GAC GCT CGG CCG GGA ACG AAT CCC ACC ACC AGG AGG AAT Asp Val Ser Asp Ala Arg Pro Gly Thr Asn Pro Thr Thr Arg Arg Asn 1720 1725 1730 1735	5657
GCC AAG GCT GGG CCC ACA GCG AGA AAC CGC TAT GCC AGC CAG TGG ACC Ala Lys Ala Gly Pro Thr Ala Arg Asn Arg Tyr Ala Ser Gln Trp Thr 1740 1745 1750	5705
CTC AAC CGA CCC CAC CCC ACC ATC TCA GCA CAC ACC CTC ACC ACA GAC Leu Asn Arg Pro His Pro Thr Ile Ser Ala His Thr Leu Thr Thr Asp 1755 1760 1765	5753
TGG AGG CTG CCA ACA CCC AGG GCT GCA GGA TCA GTA GAC AAA GAG AGC Trp Arg Leu Pro Thr Pro Arg Ala Ala Gly Ser Val Asp Lys Glu Ser 1770 1775 1780	5801
GAC AGT TAC AGC GTC AGC CCC TCG CAA GAC ACA GAT CGA GCA AGA AGC Asp Ser Tyr Ser Val Ser Pro Ser Gln Asp Thr Asp Arg Ala Arg Ser 1785 1790 1795	5849
AGC ATG GTC TCC ACA GAA AGT GCC TCC TCC ACT TAC GAA GAA CTG GCC Ser Met Val Ser Thr Glu Ser Ala Ser Ser Thr Tyr Glu Glu Leu Ala 1800 1805 1810 1815	5897
AGG GCC TAC GAA CAC GCC AAG ATG GAA GAG CAA CTG AGG CAC GCC AAG Arg Ala Tyr Glu His Ala Lys Met Glu Glu Gln Leu Arg His Ala Lys 1820 1825 1830	5945
TTC ACC ATC ACG GAG TGC TTC ATA TCA GAC ACG TCA TCG GAG CAG TTG Phe Thr Ile Thr Glu Cys Phe Ile Ser Asp Thr Ser Ser Glu Gln Leu 1835 1840 1845	5993
ACG GCA GGG ACA AAT GAG TAC ACG GAC AGT CTG ACC TCC AGC ACC CCT Thr Ala Gly Thr Asn Glu Tyr Thr Asp Ser Leu Thr Ser Ser Thr Pro 1850 1855 1860	6041
TCC GAA TCG GGA ATC TGC AGG TTC ACT GCA TCT CCC CCC AAA CCT CAG Ser Glu Ser Gly Ile Cys Arg Phe Thr Ala Ser Pro Pro Lys Pro Gln 1865 1870 1875	6089
GAT GGA GGA AGA GTA ATG AAT ATG GCA GTT CCA AAG GCA ATC GGC CAG Asp Gly Gly Arg Val Met Asn Met Ala Val Pro Lys Ala Ile Gly Gln 1880 1885 1890 1895	6137
GTG ACC TCA TAC ATT TGC CTC CAT ACC TTA GAA TGG ACT TTT TGT TAAACCGAGG Val Thr Ser Tyr Ile Cys Leu His Thr Leu Glu Trp Thr Phe Cys 1900 1905 1910	
TGGTCCAGGC ACCAGCAGGG ACCTGAGCTT AGGACAAGCA TGCTTGGAAC CTCAGAAAAG	6252
CCGGACCCTG AAGCGCCCCA CGGTCCTGGA GCCCATCCCG ATGGAAGCCG CCTCCTCCGC	6312
CTCCTCCACG AGAGAAGGAC AGTCGTGGCA GCCGGGGGCC GTGGCCACAT TACCTCAGCG	6372
GGAGGGAGCA GAGCTGGGAC AGGCAGCTAA AATGAGCAGC TCCCAAGAAT CACTGCTCGA	6432
CTCCCGGGGC CATTTGAAAG GAAACAATCC TTACGCAAAA TCTTACACCC TGGTATAACA	6492
GACAGCATGA CTGGACAGCG GTTGTAATA CAATTCAAAC AATTCAATCA AAGCTACCTT	6552
TTTTTTACGG AATTCCAATA TTTATAATTA AAGAAAATTG CAAAATATA TT	6604

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1910 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Met Trp Ile Leu Ala Leu Ser Leu Phe Gln Ser Phe Ala Asn Val Phe
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Ser Glu Asp Leu His Ser Ser Leu Tyr Phe Val Asn Ala Ser Leu Gln
          20           25           30
Glu Val Val Phe Ala Ser Thr Thr Gly Thr Leu Val Pro Cys Pro Ala
          35           40           45
Ala Gly Ile Pro Pro Val Thr Leu Arg Trp Tyr Leu Ala Thr Gly Glu
          50           55           60
Glu Ile Tyr Asp Val Pro Gly Ile Arg His Val His Pro Asn Gly Thr
          65           70           75           80
Leu Gln Ile Phe Pro Phe Pro Pro Ser Ser Phe Ser Thr Leu Ile His
          85           90           95
Asp Asn Thr Tyr Tyr Cys Thr Ala Glu Asn Pro Ser Gly Lys Ile Arg
          100          105          110
Ser Gln Asp Val His Ile Lys Ala Val Leu Arg Glu Pro Tyr Thr Val
          115          120          125
Arg Val Glu Asp Gln Lys Thr Met Arg Gly Asn Val Ala Val Phe Lys
          130          135          140
Cys Ile Ile Pro Ser Ser Val Glu Ala Tyr Ile Thr Val Val Ser Trp
          145          150          155          160
Glu Lys Asp Thr Val Ser Leu Val Ser Gly Ser Arg Phe Leu Ile Thr
          165          170          175
Ser Thr Gly Ala Leu Tyr Ile Lys Asp Val Gln Asn Glu Asp Gly Leu
          180          185          190
Tyr Asn Tyr Arg Cys Ile Thr Arg His Arg Tyr Thr Gly Glu Thr Arg
          195          200          205
Gln Ser Asn Ser Ala Arg Leu Phe Val Ser Asp Pro Ala Asn Ser Ala
          210          215          220
Pro Ser Ile Leu Asp Gly Phe Asp His Arg Lys Ala Met Ala Gly Gln
          225          230          235          240
Arg Val Glu Leu Pro Cys Lys Ala Leu Gly His Pro Glu Pro Asp Tyr
          245          250          255
Arg Trp Leu Lys Asp Asn Met Pro Leu Glu Leu Ser Gly Arg Phe Gln
          260          265          270

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009956591.10E9

[illegible]

Ala 945	Thr	Ile	Ile	Asp 950	Ile	His	Pro	Ser	Ser	Thr 955	Tyr	Ser	Ile	Arg	Met 960
Tyr	Ala	Lys	Asn 965	Arg	Ile	Gly	Lys	Ser	Glu 970	Pro	Ser	Asn	Glu	Leu 975	Thr
Ile	Thr	Ala	Asp 980	Glu	Ala	Ala	Pro	Asp 985	Gly	Pro	Pro	Gln	Glu	Val 990	His
Leu	Glu	Pro 995	Ile	Ser	Ser	Gln	Ser 1000	Ile	Arg	Val	Thr	Trp 1005	Lys	Ala	Pro
Lys	Lys 1010	His	Leu	Gln	Asn	Gly 1015	Ile	Ile	Arg	Gly	Tyr 1020	Gln	Ile	Gly	Tyr
Arg 1025	Glu	Tyr	Ser	Thr	Gly 1030	Gly	Asn	Phe	Gln	Phe 1035	Asn	Ile	Ile	Ser	Val 1040
Asp	Thr	Ser	Gly 1045	Asp	Ser	Glu	Val	Tyr 1050	Thr	Leu	Asp	Asn	Leu	Asn 1055	Lys
Phe	Thr	Gln	Tyr 1060	Gly	Leu	Val	Val	Gln 1065	Ala	Cys	Asn	Arg	Ala	Gly 1070	Thr
Gly	Pro 1075	Ser	Ser	Gln	Glu	Ile 1080	Ile	Thr	Thr	Thr	Leu	Glu 1085	Asp	Val	Pro
Ser 1090	Tyr	Pro	Pro	Glu	Asn	Val 1095	Gln	Ala	Ile	Ala	Thr 1100	Ser	Pro	Glu	Ser
Ile 1105	Ser	Ile	Ser	Trp	Ser 1110	Thr	Leu	Ser	Lys	Glu 1115	Ala	Leu	Asn	Gly	Ile 1120
Leu	Gln	Gly	Phe 1125	Arg	Val	Ile	Tyr	Trp	Ala 1130	Asn	Leu	Met	Asp	Gly 1135	Glu
Leu	Gly	Glu	Ile 1140	Lys	Asn	Ile	Thr	Thr 1145	Thr	Gln	Pro	Ser	Leu	Glu 1150	Leu
Asp 1155	Gly	Leu	Glu	Lys	Tyr	Thr 1160	Asn	Tyr	Ser	Ile	Gln 1165	Val	Leu	Ala	Phe
Thr 1170	Arg	Ala	Gly	Asp	Gly	Val 1175	Arg	Ser	Glu	Gln	Ile 1180	Phe	Thr	Arg	Thr
Lys 1185	Glu	Asp	Val	Pro	Gly 1190	Pro	Pro	Ala	Gly	Val 1195	Lys	Ala	Ala	Ala	Ala 1200
Ser	Ala	Ser	Met 1205	Val	Phe	Val	Ser	Trp	Leu 1210	Pro	Pro	Leu	Lys	Leu 1215	Asn
Gly	Ile	Ile	Arg 1220	Lys	Tyr	Thr	Val	Phe 1225	Cys	Ser	His	Pro	Tyr 1230	Pro	Thr
Val	Ile 1235	Ser	Glu	Phe	Glu	Ala 1240	Ser	Pro	Asp	Ser	Phe 1245	Ser	Tyr	Arg	Ile
Pro 1250	Asn	Leu	Ser	Arg	Asn	Arg 1255	Gln	Tyr	Ser	Val	Trp 1260	Val	Val	Ala	Val
Thr 1265	Ser	Ala	Gly	Arg	Gly 1270	Asn	Ser	Ser	Glu	Ile 1275	Ile	Thr	Val	Glu	Pro 1280

Leu	Ala	Lys	Ala	Pro	Ala	Arg	Ile	Leu	Thr	Phe	Ser	Gly	Thr	Val	Thr	
				1285					1290						1295	
Thr	Pro	Trp	Met	Lys	Asp	Ile	Val	Leu	Pro	Cys	Lys	Ala	Val	Gly	Asp	
			1300					1305					1310			
Pro	Ser	Pro	Ala	Val	Lys	Trp	Met	Lys	Asp	Ser	Asn	Gly	Thr	Pro	Ser	
			1315				1320					1325				
Leu	Val	Thr	Ile	Asp	Gly	Arg	Arg	Ser	Ile	Phe	Ser	Asn	Gly	Ser	Phe	
	1330					1335					1340					
Ile	Ile	Arg	Thr	Val	Lys	Ala	Glu	Asp	Ser	Gly	Tyr	Tyr	Ser	Cys	Ile	
1345					1350					1355					1360	
Ala	Asn	Asn	Asn	Trp	Gly	Ser	Asp	Glu	Ile	Ile	Leu	Asn	Leu	Gln	Val	
				1365					1370					1375		
Gln	Val	Pro	Pro	Asp	Gln	Pro	Arg	Leu	Thr	Val	Ser	Lys	Thr	Thr	Ser	
			1380					1385					1390			
Ser	Ser	Ile	Thr	Leu	Ser	Trp	Leu	Pro	Gly	Asp	Asn	Gly	Gly	Ser	Ser	
		1395					1400					1405				
Ile	Arg	Gly	Tyr	Ile	Leu	Gln	Tyr	Ser	Glu	Asp	Asn	Ser	Glu	Gln	Trp	
	1410					1415					1420					
Gly	Ser	Phe	Pro	Ile	Ser	Pro	Ser	Glu	Arg	Ser	Tyr	Arg	Leu	Glu	Asn	
1425					1430					1435					1440	
Leu	Lys	Cys	Gly	Thr	Trp	Tyr	Lys	Phe	Thr	Leu	Thr	Ala	Gln	Asn	Gly	
				1445					1450					1455		
Val	Gly	Pro	Gly	Arg	Ile	Ser	Glu	Ile	Ile	Glu	Ala	Lys	Thr	Leu	Gly	
			1460					1465					1470			
Lys	Glu	Pro	Gln	Phe	Ser	Lys	Glu	Gln	Glu	Leu	Phe	Ala	Ser	Ile	Asn	
	1475						1480					1485				
Thr	Thr	Arg	Val	Arg	Leu	Asn	Leu	Ile	Gly	Trp	Asn	Asp	Gly	Gly	Cys	
	1490					1495					1500					
Pro	Ile	Thr	Ser	Phe	Thr	Leu	Glu	Tyr	Arg	Pro	Phe	Gly	Thr	Thr	Val	
1505					1510					1515					1520	
Trp	Thr	Thr	Ala	Gln	Arg	Thr	Ser	Leu	Ser	Lys	Ser	Tyr	Ile	Leu	Tyr	
				1525					1530					1535		
Asp	Leu	Gln	Glu	Ala	Thr	Trp	Tyr	Glu	Leu	Gln	Met	Arg	Val	Cys	Asn	
			1540					1545					1550			
Ser	Ala	Gly	Cys	Ala	Glu	Lys	Gln	Ala	Asn	Phe	Ala	Thr	Leu	Asn	Tyr	
		1555					1560					1565				
Asp	Gly	Ser	Thr	Ile	Pro	Pro	Leu	Ile	Lys	Ser	Val	Val	Gln	Asn	Glu	
	1570				1575						1580					
Glu	Gly	Leu	Thr	Thr	Asn	Glu	Gly	Leu	Lys	Met	Leu	Val	Thr	Ile	Ser	
1585					1590					1595					1600	
Cys																

Arg Arg Arg Arg Arg Glu Gln Arg Leu Lys Arg Leu Arg Asp Ala Lys
 1620 1625 1630
 Ser Leu Ala Glu Met Leu Met Ser Lys Asn Thr Arg Thr Ser Asp Thr
 1635 1640 1645
 Leu Ser Lys Gln Gln Gln Thr Leu Arg Met His Ile Asp Ile Pro Arg
 1650 1655 1660
 Ala Gln Leu Leu Ile Glu Glu Arg Asp Thr Met Glu Thr Ile Asp Asp
 1665 1670 1675 1680
 Arg Ser Thr Val Leu Leu Thr Asp Ala Asp Phe Gly Glu Ala Ala Lys
 1685 1690 1695
 Gln Lys Ser Leu Thr Val Thr His Thr Val His Tyr Gln Ser Val Ser
 1700 1705 1710
 Gln Ala Thr Gly Pro Leu Val Asp Val Ser Asp Ala Arg Pro Gly Thr
 1715 1720 1725
 Asn Pro Thr Thr Arg Arg Asn Ala Lys Ala Gly Pro Thr Ala Arg Asn
 1730 1735 1740
 Arg Tyr Ala Ser Gln Trp Thr Leu Asn Arg Pro His Pro Thr Ile Ser
 1745 1750 1755 1760
 Ala His Thr Leu Thr Thr Asp Trp Arg Leu Pro Thr Pro Arg Ala Ala
 1765 1770 1775
 Gly Ser Val Asp Lys Glu Ser Asp Ser Tyr Ser Val Ser Pro Ser Gln
 1780 1785 1790
 Asp Thr Asp Arg Ala Arg Ser Ser Met Val Ser Thr Glu Ser Ala Ser
 1795 1800 1805
 Ser Thr Tyr Glu Glu Leu Ala Arg Ala Tyr Glu His Ala Lys Met Glu
 1810 1815 1820
 Glu Gln Leu Arg His Ala Lys Phe Thr Ile Thr Glu Cys Phe Ile Ser
 1825 1830 1835 1840
 Asp Thr Ser Ser Glu Gln Leu Thr Ala Gly Thr Asn Glu Tyr Thr Asp
 1845 1850 1855
 Ser Leu Thr Ser Ser Thr Pro Ser Glu Ser Gly Ile Cys Arg Phe Thr
 1860 1865 1870
 Ala Ser Pro Pro Lys Pro Gln Asp Gly Gly Arg Val Met Asn Met Ala
 1875 1880 1885
 Val Pro Lys Ala Ile Gly Gln Val Thr Ser Tyr Ile Cys Leu His Thr
 1890 1895 1900
 Leu Glu Trp Thr Phe Cys
 1905 1910

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 388 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: both
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

```

CCGGGTATTC TTACTCATGA GCATTTTCAGC TAAACTCTTT GCATCTCGCA GCCTCTTTAG      60
CCTCTGCTCC CGCCGCCTCC TCCGCACAAC CAGCAGGAGC ACAAACAGCA GCAAGACCCC      120
CACCAGGATA CAGGAGATGG TCACCAGCAT CTTGAGCCCC TCGTTGGTCG TCAGCCCTTC      180
TTCGTTTTTG ACAACTGACT TAATGAGTGG AGGAATTGTA CTGCCATCGT AGTTCAGCGT      240
AGCGAAGTTG GCCTGCTTCT CCGCGCAGCC CGCACTGTTG CACACCCGCA TCTGCAGCTC      300
ATACCAGGTG GCTTCCTGCA GGTACATACAG GATGTAGGAC TTGGAGAGAG AGGTCCTCTG      360
AGCTGTGGTC CAAACTGTGG TCCCAAAG                                         388

```

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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CCTGATGCTC GAGTGAATTC                                         20

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(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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CCAGTTCTCA AAGGAGCAGG                                         20

```

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

CCTGTATGAC CTGCAGGAAG

20

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 842 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: both
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CCGGGCCCGG CGCGGCCGGAG CGCAGCGCAA CGCGGGGGGC GAGGCCGGCG CGTGGCTCGC	60
TCGCTGGCTC GCTGGCTCGC GGGAGGCCGG GCAGCAGCAG GGGCATGTGG ATACTGGCTC	120
TCTCCTTGTT CCAGAGCTTC GCGAATGTTT TCAGTGAAGA GCCCCACTCC AGCCTCTACT	180
TTGTCAATGC ATCGCTGCAA GAGGTAGTGT TTGCAAGCAC ATCGGGGACG CTGGTGCCCT	240
GCCCGGCTGC AGGCATCCCT CCTGTGACTC TCAGATGGTA CCTAGCAACG GGCGAGGAGA	300
TCTACGATGT CCCCGGGATC CGCCACGTCC ATCCCAATGG CACTCTCCAA ATTTTCCCCT	360
TTCTCCTTC AAGCTTCAGC ACCTTAATCC ATGATAATAC TTACTATTGC ACAGCTGAAA	420
ACCCTTCAGG GAAAATTAGA AGTCAGGATG TCCACATCAA GGCTGTTTTA CGGGAGCCCT	480
ATACAGTCCG TGTGGAGGAC CAGAAAACCA TGAGAGGCAA TGTCGCGGTG TTCAAGTGCA	540
TTATCCCCTC CTCGGTGGAG GCGTACGTCT CTGTCTGCTC ATGGGAGAAA GACACGGTTT	600
CACTTGTCTC AGGATCTAGA TTTCTCATCA CATCCACGGG AGCCTTGAT ATTAAGATG	660
TTCAGAACGA AGATGGGCTG TACAACTACC GCTGCATCGC GCGGCACAGA TTCGCGGGGG	720
AGACGAGACA GAGCAACTGC GCGAGACTGT TCGTGTGAGA ACCAGCAAAC TCAGCCCATC	780
CATCCTGGAA GGGTTTGACC ACCGCCAAAC CATGGCCGGG CACGCGTGGA GCTGCCTTGC	840
CA	842

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 898 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: both
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

TGCCGGCCGG TTGCAAGCCT GTACTACAGG CCATACTGCG TGAATTATCA GGTGTGCCAG	60
GGTGTACACT TCGCTGTCCC GGTGGTGTCA ATACTGATGA TGTGAACTG GAAGTTACCC	120
CGTGCTGTAC TCCGGTAGCC TATTGGTAGC CGCGAATGAT CCCGTCTTGT ATAGTGTCTT	180
TGGGAGCCTC TCCAGGTAAC CCTGATACTC TGAGATGAGG TGGGTTCCTAA GTGAACTTCC	240
TGAGGTGGAC ATCACGAGCT GCCTCATCCG CCGTGATGGT GATCTCGTTG CTGGGCTCAC	300
TCTTGCCAAT CCGGTTCTTG GCGTACATGC GGATGCTGTA GGTGGAGGAA GGTGGATAT	360
CAATGATGGT GGCCGAGTTC AGCTGAGGGG AAACATCTTT GGTCTCTGA GCAGAATCCC	420
ACGAGTCTGA TTTATTTTTG CATTCACT GTCATAGCCT GTGATGGGGC TGTTGCCATC	480
AAACCCCATG GTCCACCTGA GCGTGATGGT GCGAGCTTTG ACATCTCTTG ATCTCAATCT	540
CGGGAGGATC TGGGGGTTCT TGCCTGTGA GTTGAATTAT TCCACGGTCC TCCCCGTATG	600
AATTGATAGC ATGGCAGGAG AAGAAACCGG AATCTTCTCT CACTGTTGGC AAAATCTGCA	660
GCGTAGATAT CACTTCCTCT CCCACCTCCT TGGTGGATAC AGTACGGGCC ACTTTCAGGG	720
TTAATGATCC TGTCTCTCTT CTCCAGCGGA CAATGATGGG CTCTCCCATG GGCTGTGCAG	780
CTCATTCCTT CCTTTGACCC TGATGGCCAG GTGGTGTGGG TATAAGTTAT ATCATGGCCC	840
GAATTTCCCT GTGAGTCCAT GGAATTGCTG AACGTCTGC GCCACATCG TTCGCTGA	898

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2173 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: both
 (D) TOPOLOGY: both

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ACCACCATTC ACACACCCAG ACATGGCGGG TCGCGGCAA CCTTCAGTTC CTGGCCTTCC	60
TGTAGGGTAA AGGGCTGCTG CGGGTTTATA GACCGGCACA TGCCCATCCT GGCATACGGT	120
GGCCAGTGGC TTTCCATCTG GATTCCAGGC CAAGCTAAAA ATCTGTTCTT GATGGCCCTG	180

(2) INFORMATION FOR SEQ ID NO:10:

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TGACTGAGGC	CGGAGCACGG	CAAAGATGAG	CCTGCCCCGCC	CGCCTGCTGC	CTGGATGCGG	60										
AGGGTGAGGG	CTGGCGCACG	GGAGGCCCGCT	GGTGTCGCAT	TCTGGGCGCC	GAGTGCCCCG	120										
GATGAGCTCA	CGCCCGCGTC	TGCGGCTCTC	TCCACCTGCC	GACCTGCCGG	GGGCCCCACTG	180										
AGCTGACGGC	GCACCTGGGC	TCCGGCCGCA	GCGTGGGGCG	CGGCGCCCCG	GAGCAGGTGT	240										
GCAGGAGCGC	AGCGCGCGGC	GAGCGCAGCC	CTCGCTCCGG	AGCCCGGCCG	CGCCGCGTGC	300										
CCGGGCGGCT	AGGCAGCGGC	GGCGGCGGCG	GCGGGCGGCG	GGCGGGCGGC	GGCCCCCGGG	360										
CAGGTGCCGA	GCGGCGAGCG	GAGCCGGGCC	GGGCGGAGCG	CGGGGGGCGA	GGCCGGCGCG	420										
TCGCTCGCGG	GAGGCCGGGG	AGCGGCAGGG	GC	ATG	TGG	ATA	CTG	GCT	CTC	TCC		473				
			Met	Trp	Ile	Leu	Ala	Leu	Ser							
			1					5								
TTG	TTC	CAG	AGC	TTC	GCG	AAT	GTT	TTC	AGT	GAA	GAC	CTA	CAC	TCC	AGC	521
Leu	Phe	Gln	Ser	Phe	Ala	Asn	Val	Phe	Ser	Glu	Asp	Leu	His	Ser	Ser	
		10					15					20				
CTC	TAC	TTT	GTC	AAT	GCA	TCT	CTG	CAA	GAG	GTA	GTG	TTT	GCC	AGC	ACC	569
Leu	Tyr	Phe	Val	Asn	Ala	Ser	Leu	Gln	Glu	Val	Val	Phe	Ala	Ser	Thr	
		25				30					35					
ACG	GGG	ACT	CTG	GTG	CCC	TGC	CCC	GCA	GCA	GGC	ATC	CCT	CCT	GTG	ACT	617
Thr	Gly	Thr	Leu	Val	Pro	Cys	Pro	Ala	Ala	Gly	Ile	Pro	Pro	Val	Thr	
		40			45					50					55	
CTC	AGA	TGG	TAC	CTA	GCC	ACG	GGC	GAG	GAG	ATC	TAC	GAT	GTC	CCC	GGG	665
Leu	Arg	Trp	Tyr	Leu	Ala	Thr	Gly	Glu	Glu	Ile	Tyr	Asp	Val	Pro	Gly	
				60				65						70		
ATC	CGC	CAC	GTC	CAC	CCC	AAC	GGC	ACT	CTC	CAA	ATT	TTC	CCC	TTC	CCT	713
Ile	Arg	His	Val	His	Pro	Asn	Gly	Thr	Leu	Gln	Ile	Phe	Pro	Phe	Pro	
			75					80					85			
CCT	TCA	AGC	TTC	AGT	ACC	TTA	ATC	CAT	GAT	AAT	ACT	TAT	TAT	TGC	ACA	761
Pro	Ser	Ser	Phe	Ser	Thr	Leu	Ile	His	Asp	Asn	Thr	Tyr	Tyr	Cys	Thr	
		90					95					100				

GCT Ala	GAA Glu	AAT Asn	CCT Pro	TCA Ser	GGG Gly	AAA Lys	ATT Ile	AGA Arg	AGT Ser	CAG Gln	GAT Asp	GTC Val	CAC His	ATC Ile	AAG Lys	809
105110115																
GCT Ala	GTT Val	TTA Leu	CGG Arg	GAG Glu	CCC Pro	TAT Tyr	ACA Thr	GTC Val	CGT Arg	GTG Val	GAG Glu	GAC Asp	CAG Gln	AAA Lys	ACC Thr	857
120125130135																
ATG Met	AGA Arg	GGC Gly	AAT Asn	GTT Val	GCG Ala	GTC Val	TTC Phe	AAG Lys	TGC Cys	ATT Ile	ATC Ile	CCC Pro	TCC Ser	TCG Ser	GTG Val	905
140145150																
GAG Glu	GCG Ala	TAC Tyr	ATC Ile	ACT Thr	GTC Val	GTC Val	TCA Ser	TGG Trp	GAG Glu	AAA Lys	GAC Asp	ACT Thr	GTT Val	TCA Ser	CTT Leu	953
155160165																
GTC Val	TCA Ser	GGA Gly	TCT Ser	AGA Arg	TTT Phe	CTC Leu	ATC Ile	ACA Thr	TCC Ser	ACG Thr	GGA Gly	GCC Ala	TTG Leu	TAT Tyr	ATT Ile	1001
170175180																
AAA Lys	GAT Asp	GTA Val	CAG Gln	AAT Asn	GAA Glu	GAT Asp	GGA Gly	TTG Leu	TAT Tyr	AAC Asn	TAC Tyr	CGC Arg	TGC Cys	ATC Ile	ACG Thr	1049
185190195																
CGG Arg	CAT His	CGA Arg	TAC Tyr	ACC Thr	GGA Gly	GAG Glu	ACG Thr	AGG Arg	CAG Gln	AGC Ser	AAC Asn	AGC Ser	GCC Ala	AGA Arg	CTT Leu	1097
200205210215																
TTT Phe	GTA Val	TCA Ser	GAC Asp	CCA Pro	GCG Ala	AAC Asn	TCA Ser	GCC Ala	CCA Pro	TCC Ser	ATA Ile	CTG Leu	GAT Asp	GGG Gly	TTT Phe	1145
220225230																
GAC Asp	CAT His	CGC Arg	AAA Lys	GCC Ala	ATG Met	GCT Ala	GGG Gly	CAG Gln	CGT Arg	GTG Val	GAG Glu	CTG Leu	CCT Pro	TGC Cys	AAA Lys	1193
235240245																
GCG Ala	CTC Leu	GGG Gly	CAC His	CCT Pro	GAG Glu	CCA Pro	GAT Asp	TAC Tyr	CGC Arg	TGG Trp	CTG Leu	AAG Lys	GAC Asp	AAC Asn	ATG Met	1241
250255260																
CCC Pro	CTG Leu	GAA Glu	CTT Leu	TCA Ser	GGG Gly	AGG Arg	TTC Phe	CAG Gln	AAG Lys	ACC Thr	GTG Val	ACG Thr	GGG Gly	CTG Leu	CTC Leu	1289
265270275																
ATT Ile	GAG Glu	AAC Asn	ATT Ile	CGC Arg	CCC Pro	TCG Ser	GAC Asp	TCA Ser	GGC Gly	AGC Ser	TAT Tyr	GTT Val	TGT Cys	GAA Glu	GTG Val	1337
280285290295																
TCC Ser	AAC Asn	AGA Arg	TAC Tyr	GGA Gly	ACT Thr	GCT Ala	AAG Lys	GTG Val	ATA Ile	GGC Gly	CGC Arg	CTG Leu	TAC Tyr	GTG Val	AAA Lys	1385
300305310																
CAG Gln	CCA Pro	CTG Leu	AAA Lys	GCC Ala	ACC Thr	ATC Ile	AGT Ser	CCC Pro	AGG Arg	AAG Lys	GTT Val	AAA Lys	AGC Ser	AGC Ser	GTG Val	1433
315320325																
GGT Gly	AGC Ser	CAA Gln	GTT Val	TCC Ser	TTG Leu	TCC Ser	TGC Cys	AGC Ser	GTG Val	ACA Thr	GGA Gly	ACT Thr	GAG Glu	GAC Asp	CAG Gln	1481
330335340																
GAA Glu	CTC Leu	TCC Ser	TGG Trp	TAC Tyr	CGC Arg	AAT Asn	GGT Gly	GAA Glu	ATC Ile	CTC Leu	AAC Asn	CCT Pro	GGA Gly	AAA Lys	AAT Asn	1529
345350355																

GTG Val 360	AGG Arg	ATC Ile	ACA Thr	GGG Gly 365	ATC Ile 365	AAC Asn	CAC His	GAA Glu	AAC Asn	CTT Leu 370	ATA Ile	ATG Met	GAT Asp	CAC His	ATG Met 375	1577
GTC Val	AAA Lys	AGT Ser	GAC Asp	GGG Gly 380	GGC Gly	GCA Ala	TAC Tyr	CAG Gln	TGC Cys 385	TTT Phe	GTG Val	CGC Arg	AAG Lys	GAC Asp 390	AAG Lys	1625
CTG Leu	TCC Ser	GCT Ala	CAA Gln 395	GAC Asp	TAT Tyr	GTG Val	CAG Gln	GTG Val 400	GTC Val	CTT Leu	GAA Glu	GAT Asp	GGG Gly 405	ACT Thr	CCC Pro	1673
AAA Lys	ATT Ile	ATT Ile 410	TCT Ser	GCC Ala	TTT Phe	AGT Ser	GAA Glu 415	AAG Lys	GTG Val	GTG Val	AGT Ser	CCA Pro 420	GCA Ala	GAG Glu	CCG Pro	1721
GTT Val	TCC Ser 425	CTT Leu	ATG Met	TGC Cys	AAC Asn	GTG Val 430	AAG Lys	GGA Gly	ACA Thr	CCT Pro	TTG Leu 435	CCC Pro	ACG Thr	ATC Ile	ACG Thr	1769
TGG Trp 440	ACC Thr	CTG Leu	GAC Asp	GAT Asp 445	GAC Asp	CCG Pro	ATT Ile	CTC Leu	AAG Lys	GGT Gly 450	GGC Gly	AGT Ser	CAC His	CGC Arg	ATC Ile 455	1817
AGC Ser	CAG Gln	ATG Met	ATC Ile 460	ACG Thr	TCG Ser	GAG Glu	GGG Gly	AAC Asn 465	GTG Val	GTC Val	AGC Ser	TAC Tyr	CTG Leu	AAC Asn 470	ATC Ile	1865
TCC Ser	AGC Ser	TCC Ser	CAG Gln 475	GTC Val	CGG Arg	GAC Asp	GGG Gly	GGA Gly 480	GTC Val	TAC Tyr	CGC Arg	TGC Cys	ACT Thr 485	GCC Ala	AAC Asn	1913
AAC Asn	TCG Ser 490	GCG Ala	GGA Gly	GTC Val	GTC Val	CTG Leu	TAC Tyr 495	CAG Gln	GCT Ala	CGA Arg	ATA Ile	AAC Asn 500	GTA Val	AGA Arg	GGG Gly	1961
CCT Pro	GCA Ala 505	AGC Ser	ATT Ile	CGA Arg	CCA Pro	ATG Met 510	AAA Lys	AAC Asn	ATC Ile	ACA Thr	GCA Ala 515	ATA Ile	GCA Ala	GGA Gly	CGG Arg	2009
GAC Asp 520	ACA Thr	TAC Tyr	ATT Ile	CAC His	TGT Cys 525	CGT Arg	GTG Val	ATT Ile	GGC Gly	TAT Tyr 530	CCG Pro	TAT Tyr	TAC Tyr	TCC Ser	ATT Ile 535	2057
AAA Lys	TGG Trp	TAC Tyr	AAG Lys	AAC Asn 540	TCT Ser	AAC Asn	CTG Leu	CTT Leu	CCT Pro 545	TTC Phe	AAC Asn	CAC His	CGC Arg	CAA Gln 550	GTG Val	2105
GCA Ala	TTT Phe	GAG Glu	AAC Asn 555	AAT Asn	GGA Gly	ACT Thr	CTT Leu	AAA Lys 560	CTT Leu	TCA Ser	GAT Asp	GTG Val	CAA Gln 565	AAG Lys	GAA Glu	2153
GTG Val	GAC Asp	GAG Glu	GGG Gly 570	GAG Glu	TAC Tyr	ACG Thr	TGC Cys 575	AAC Asn	GTG Val	TTG Leu	GTT Val	CAA Gln	CCA Pro	CAA Gln	CTC Leu	2201
TCC Ser	ACC Thr 585	AGC Ser	CAG Gln	AGC Ser	GTC Val	CAC His 590	GTG Val	ACC Thr	GTG Val	AAA Lys	GTT Val	CCG Pro	CCT Pro	TTC Phe	ATA Ile	2249
CAA Gln 600	CCC Pro	TTT Phe	GAG Glu	TTT Phe	CCA Pro 605	AGA Arg	TTC Phe	TCC Ser	ATT Ile	GGG Gly 610	CAG Gln	CGG Arg	GTC Val	TTC Phe	ATC Ile 615	2297

CCC Pro	TGT Cys	GTT Val	GTG Val	GTC Val 620	TCA Ser	GGG Gly	GAC Asp	TTA Leu	CCC Pro 625	ATC Ile	ACG Thr	ATC Ile	ACC Thr	TGG Trp 630	CAG Gln	2345
AAG Lys	GAT Asp	GGC Gly	CGG Arg 635	CCA Pro	ATC Ile	CCT Pro	GGG Gly 640	AGC Ser	CTT Leu	GGG Gly	GTG Val	ACC Thr	ATT Ile 645	GAC Asp	AAT Asn	2393
ATT Ile	GAC Asp	TTC Phe 650	ACG Thr	AGC Ser	TCC Ser	TTG Leu	AGG Arg 655	ATT Ile	TCC Ser	AAT Asn	CTC Leu	TCG Ser 660	CTC Leu	ATG Met	CAC His	2441
AAT Asn	GGG Gly 665	AAT Asn	TAC Tyr	ACC Thr	TGC Cys	ATA Ile 670	GCC Ala	CGG Arg	AAT Asn	GAG Glu	GCC Ala 675	GCC Ala	GCT Ala	GTG Val	GAG Glu	2489
CAC His 680	CAA Gln	AGC Ser	CAG Gln	TTG Leu 685	ATT Ile	GTC Val	AGA Arg	GTT Val	CCT Pro	CCC Pro 690	AAG Lys	TTT Phe	GTG Val	GTT Val	CAG Gln 695	2537
CCA Pro	CGG Arg	GAC Asp	CAG Gln 700	GAC Asp	GGG Gly	ATT Ile	TAT Tyr	GGC Gly	AAA Lys 705	GCA Ala	GTC Val	ATC Ile	CTC Leu	AAT Asn 710	TGT Cys	2585
TCT Ser	GCT Ala	GAG Glu	GGT Gly 715	TAC Tyr	CCT Pro	GTA Val	CCT Pro	ACC Thr 720	ATC Ile	GTG Val	TGG Trp	AAA Lys	TTC Phe 725	TCT Ser	AAA Lys	2633
GGT Gly	GCT Ala	GGG Gly 730	GTT Val	CCC Pro	CAG Gln	TTC Phe	CAG Gln 735	CCA Pro	ATT Ile	GCC Ala	CTA Leu	AAT Asn 740	GGC Gly	CGA Arg	ATC Ile	2681
CAA Gln 745	GTT Val	CTC Leu	AGC Ser	AAT Asn	GGG Gly	TCG Ser 750	TTG Leu	CTG Leu	ATC Ile	AAG Lys	CAT His 755	GTC Val	GTG Val	GAG Glu	GAA Glu	2729
GAC Asp 760	AGT Ser	GGC Gly	TAC Tyr	TAC Tyr	CTC Leu 765	TGC Cys	AAG Lys	GTC Val	AGC Ser	AAC Asn 770	GAT Asp	GTG Val	GGC Gly	GCA Ala	GAC Asp 775	2777
GTC Val	AGC Ser	AAG Lys	TCC Ser	ATG Met 780	TAC Tyr	CTC Leu	ACG Thr	GTT Val	AAA Lys 785	ATT Ile	CCT Pro	GCG Ala	ATG Met	ATA Ile 790	ACA Thr	2825
TCC Ser	TAT Tyr	CCA Pro	AAT Asn 795	ACT Thr	ACC Thr	CTG Leu	GCC Ala	ACG Thr 800	CAG Gln	GGG Gly	CAG Gln	AAA Lys	AAG Lys 805	GAG Glu	ATG Met	2873
AGC Ser	TGC Cys	ACG Thr 810	GCG Ala	CAT His	GGT Gly	GAG Glu	AAG Lys 815	CCC Pro	ATT Ile	ATA Ile	GTC Val	CGC Arg 820	TGG Trp	GAG Glu	AAG Lys	2921
GAG Glu	GAC Asp 825	CGA Arg	ATC Ile	ATT Ile	AAC Asn	CCT Pro 830	GAG Glu	ATG Met	GCC Ala	CGT Arg	TAT Tyr 835	CTT Leu	GTG Val	TCC Ser	ACC Thr	2969
AAG Lys 840	GAG Glu	GTG Val	GGA Gly	GAA Glu 845	GAG Glu	GTG Val	ATT Ile	TCT Ser	ACT Thr	CTG Leu 850	CAG Gln	ATT Ile	TTG Leu	CCA Pro	ACT Thr 855	3017
GTG Val	AGA Arg	GAA Glu	GAT Asp 860	TCT Ser	GGT Gly	TTC Phe	TTT Phe	TCC Ser	TGC Cys 865	CAT His	GCT Ala	ATT Ile	AAT Asn	TCT Ser 870	TAT Tyr	3065

GGG Gly	GAG Glu	GAC Asp	CGT Arg 875	GGA Gly	ATA Ile	ATT Ile	CAG Gln	CTC Leu 880	ACA Thr	GTG Val	CAA Gln	GAG Glu	CCC Pro 885	CCA Pro	GAC Asp	3113
CCT Pro	CCC Pro	GAA Glu 890	ATT Ile	GAG Glu	ATC Ile	AAA Lys	GAT Asp 895	GTC Val	AAA Lys	GCA Ala	CGC Arg	ACA Thr 900	ATT Ile	ACG Thr	CTC Leu	3161
AGG Arg	TGG Trp 905	ACC Thr	ATG Met	GGG Gly	TTT Phe	GAT Asp 910	GGA Gly	AAC Asn	AGT Ser	CCC Pro	ATC Ile 915	ACA Thr	GGC Gly	TAC Tyr	GAT Asp	3209
ATT Ile 920	GAA Glu	TGC Cys	AAA Lys	AAT Asn	AAA Lys 925	TCA Ser	GAC Asp	TCC Ser	TGG Trp	GAT Asp 930	TCT Ser	GCT Ala	CAG Gln	AGA Arg	ACC Thr 935	3257
AAA Lys	GAT Asp	GTT Val	TCC Ser	CCT Pro 940	CAG Gln	CTG Leu	AAC Asn	TCG Ser	GCC Ala 945	ACC Thr	ATC Ile	ATT Ile	GAT Asp	ATC Ile 950	CAC His	3305
CCT Pro	TCC Ser	TCC Ser	ACC Thr 955	TAC Tyr	AGC Ser	ATC Ile	CGC Arg	ATG Met 960	TAC Tyr	GCC Ala	AAG Lys	AAC Asn	CGG Arg 965	ATT Ile	GGC Gly	3353
AAG Lys	AGC Ser	GAG Glu 970	CCC Pro	AGC Ser	AAC Asn	GAG Glu	CTC Leu 975	ACC Thr	ATC Ile	ACG Thr	GCG Ala	GAC Asp 980	GAG Glu	GCA Ala	GCT Ala	3401
CCT Pro 985	GAT Asp	GGT Gly	CCA Pro	CCT Pro	CAG Gln	GAA Glu 990	GTT Val	CAC His	CTG Leu	GAG Glu	CCT Pro 995	ATA Ile	TCA Ser	TCT Ser	CAG Gln	3449
AGC Ser 1000	ATC Ile	AGG Arg	GTC Val	ACA Thr	TGG Trp 1005	AAG Lys	GCT Ala	CCC Pro	AAG Lys	AAA Lys 1010	CAT His	TTG Leu	CAA Gln	AAT Asn	GGG Gly 1015	3497
ATT Ile	ATC Ile	CGT Arg	GGC Gly 1020	TAC Tyr	CAA Gln	ATA Ile	GGT Gly	TAC Tyr	CGA Arg 1025	GAG Glu	TAC Tyr	AGC Ser	ACT Thr	GGG Gly 1030	GGT Gly	3545
AAC Asn	TTC Phe	CAA Gln	TTC Phe 1035	AAC Asn	ATT Ile	ATC Ile	AGT Ser	GTC Val 1040	GAC Asp	ACC Thr	AGC Ser	GGG Gly	GAC Asp 1045	AGT Ser	GAG Glu	3593
GTT Val	TAC Tyr	ACC Thr 1050	CTG Leu	GAC Asp	AAC Asn	CTG Leu	AAT Asn 1055	AAG Lys	TTC Phe	ACT Thr	CAG Gln	TAC Tyr 1060	GGC Gly	CTG Leu	GTG Val	3641
GTG Val 1065	CAG Gln	GCC Ala	TGT Cys	AAC Asn	CGG Arg	GCC Ala 1070	GGC Gly	ACG Thr	GGG Gly	CCT Pro	TCT Ser 1075	TCT Ser	CAG Gln	GAA Glu	ATC Ile	3689
ATC Ile 1080	ACC Thr	ACC Thr	ACT Thr	CTC Leu	GAG Glu	GAT Asp 1085	GTG Val	CCC Pro	AGT Ser	TAC Tyr 1090	CCC Pro	CCC Pro	GAA Glu	AAT Asn	GTC Val 1095	3737
CAA Gln	GCC Ala	ATA Ile	GCA Ala 1100	ACA Thr	TCA Ser	CCA Pro	GAA Glu	AGC Ser	ATA Ile 1105	TCA Ser	ATA Ile	TCC Ser	TGG Trp	TCC Ser 1110	ACA Thr	3785
CTT Leu	TCC Ser	AAG Lys 1115	GAA Glu	GCC Ala	TTG Leu	AAT Asn	GGA Gly	ATT Ile 1120	CTC Leu	CAG Gln	GGG Gly	TTC Phe	AGA Arg 1125	GTC Val	ATT Ile	3833

TAC	TGG	GCC	AAC	CTC	ATG	GAC	GGA	GAG	CTG	GGT	GAG	ATT	AAA	AAC	ATC	3881
Tyr	Trp	Ala	Asn	Leu	Met	Asp	Gly	Glu	Leu	Gly	Glu	Ile	Lys	Asn	Ile	
		1130					1135					1140				
ACC	ACC	ACA	CAG	CCT	TCA	CTG	GAG	CTG	GAC	GGG	CTG	GAA	AAG	TAC	ACC	3929
Thr	Thr	Thr	Gln	Pro	Ser	Leu	Glu	Leu	Asp	Gly	Leu	Glu	Lys	Tyr	Thr	
		1145				1150				1155						
AAC	TAC	AGC	ATC	CAG	GTG	CTG	GCC	TTC	ACC	CGC	GCA	GGA	GAC	GGG	GTC	3977
Asn	Tyr	Ser	Ile	Gln	Val	Leu	Ala	Phe	Thr	Arg	Ala	Gly	Asp	Gly	Val	
1160					1165					1170					1175	
AGG	AGT	GAG	CAG	ATC	TTC	ACC	CGG	ACC	AAA	GAG	GAT	GTT	CCA	GGT	CCT	4025
Arg	Ser	Glu	Gln	Ile	Phe	Thr	Arg	Thr	Lys	Glu	Asp	Val	Pro	Gly	Pro	
				1180					1185					1190		
CCC	GCG	GGT	GTG	AAG	GCA	GCG	GCG	GCC	TCA	GCC	TCC	ATG	GTC	TTT	GTG	4073
Pro	Ala	Gly	Val	Lys	Ala	Ala	Ala	Ala	Ser	Ala	Ser	Met	Val	Phe	Val	
			1195					1200					1205			
TCC	TGG	CTT	CCC	CCT	CTC	AAG	CTG	AAC	GGC	ATC	ATC	CGA	AAG	TAC	ACT	4121
Ser	Trp	Leu	Pro	Pro	Leu	Lys	Leu	Asn	Gly	Ile	Ile	Arg	Lys	Tyr	Thr	
		1210				1215						1220				
GTA	TTC	TGC	TCC	CAC	CCC	TAT	CCC	ACA	GTG	ATC	AGC	GAG	TTT	GAG	GCC	4169
Val	Phe	Cys	Ser	His	Pro	Tyr	Pro	Thr	Val	Ile	Ser	Glu	Phe	Glu	Ala	
		1225				1230					1235					
TCT	CCC	GAC	TCG	TTT	TCC	TAC	AGA	ATT	CCC	AAC	CTG	AGT	AGG	AAT	CGT	4217
Ser	Pro	Asp	Ser	Phe	Ser	Tyr	Arg	Ile	Pro	Asn	Leu	Ser	Arg	Asn	Arg	
1240					1245					1250					1255	
CAG	TAC	AGC	GTC	TGG	GTG	GTG	GCT	GTT	ACT	TCA	GCC	GGA	AGA	GGC	AAC	4265
Gln	Tyr	Ser	Val	Trp	Val	Val	Ala	Val	Thr	Ser	Ala	Gly	Arg	Gly	Asn	
				1260					1265					1270		
AGC	AGT	GAA	ATC	ATC	ACA	GTC	GAG	CCA	CTA	GCA	AAA	GCT	CCT	GCA	CGA	4313
Ser	Ser	Glu	Ile	Ile	Thr	Val	Glu	Pro	Leu	Ala	Lys	Ala	Pro	Ala	Arg	
			1275					1280					1285			
ATC	CTG	ACC	TTC	AGT	GGG	ACA	GTG	ACT	ACT	CCA	TGG	ATG	AAA	GAC	ATT	4361
Ile	Leu	Thr	Phe	Ser	Gly	Thr	Val	Thr	Thr	Pro	Trp	Met	Lys	Asp	Ile	
		1290				1295						1300				
GTC	TTG	CCT	TGT	AAG	GCT	GTT	GGG	GAC	CCT	TCT	CCT	GCA	GTC	AAA	TGG	4409
Val	Leu	Pro	Cys	Lys	Ala	Val	Gly	Asp	Pro	Ser	Pro	Ala	Val	Lys	Trp	
		1305				1310						1315				
ATG	AAA	GAC	AGT	AAC	GGG	ACA	CCC	AGT	CTA	GTA	ACG	ATT	GAT	GGG	CGG	4457
Met	Lys	Asp	Ser	Asn	Gly	Thr	Pro	Ser	Leu	Val	Thr	Ile	Asp	Gly	Arg	
1320					1325					1330					1335	
AGG	AGC	ATC	TTT	AGC	AAC	GGA	AGC	TTC	ATT	ATT	CGC	ACG	GTG	AAA	GCA	4505
Arg	Ser	Ile	Phe	Ser	Asn	Gly	Ser	Phe	Ile	Ile	Arg	Thr	Val	Lys	Ala	
				1340					1345					1350		
GAA	GAC	TCC	GGC	TAT	TAC	AGC	TGC	ATT	GCC	AAT	AAC	AAC	TGG	GGA	TCT	4553
Glu	Asp	Ser	Gly	Tyr	Tyr	Ser	Cys	Ile	Ala	Asn	Asn	Asn	Trp	Gly	Ser	
			1355					1360					1365			
GAT	GAA	ATT	ATT	TTA	AAC	TTA	CAA	GTA	CAA	GTT	CCA	CCA	GAT	CAG	CCT	4601
Asp	Glu	Ile	Ile	Leu	Asn	Leu	Gln	Val	Gln	Val	Pro	Pro	Asp	Gln	Pro	
		1370					1375					1380				

CGG CTT ACA GTC TCC AAG ACC ACG TCT TCC TCC ATC ACC CTT TCT TGG 4649
 Arg Leu Thr Val Ser Lys Thr Thr Ser Ser Ser Ile Thr Leu Ser Trp
 1385 1390 1395

CTC CCT GGA GAC AAC GGG GGC AGC TCT ATC AGA GGA TAC ATA CTG CAG 4697
 Leu Pro Gly Asp Asn Gly Gly Ser Ser Ile Arg Gly Tyr Ile Leu Gln
 1400 1405 1410 1415

TAC TCC GAG GAC AAT AGT GAG CAG TGG GGG AGT TTT CCA ATC AGC CCC 4745
 Tyr Ser Glu Asp Asn Ser Glu Gln Trp Gly Ser Phe Pro Ile Ser Pro
 1420 1425 1430

AGC GAA CGT TCC TAT CGC TTG GAA AAT CTC AAA TGT GGG ACT TGG TAT 4793
 Ser Glu Arg Ser Tyr Arg Leu Glu Asn Leu Lys Cys Gly Thr Trp Tyr
 1435 1440 1445

AAG TTC ACA CTG ACA GCC CAA AAT GGA GTG GGC CCA GGG CGC ATA AGT 4841
 Lys Phe Thr Leu Thr Ala Gln Asn Gly Val Gly Pro Gly Arg Ile Ser
 1450 1455 1460

GAA ATC ATA GAA GCA AAG ACC TTA GGA AAA GAG CCC CAG TTC TCA AAG 4889
 Glu Ile Ile Glu Ala Lys Thr Leu Gly Lys Glu Pro Gln Phe Ser Lys
 1465 1470 1475

GAG CAG GAG CTG TTT GCC AGC ATC AAC ACC ACA CGC GTG AGG CTG AAC 4937
 Glu Gln Glu Leu Phe Ala Ser Ile Asn Thr Thr Arg Val Arg Leu Asn
 1480 1485 1490 1495

CTC ATT GGC TGG AAT GAT GGC GGC TGC CCC ATC ACC TCC TTC ACA CTA 4985
 Leu Ile Gly Trp Asn Asp Gly Gly Cys Pro Ile Thr Ser Phe Thr Leu
 1500 1505 1510

GAG TAC AGG CCC TTT GGG ACC ACA GTT TGG ACC ACA GCT CAG AGG ACC 5033
 Glu Tyr Arg Pro Phe Gly Thr Thr Val Trp Thr Thr Ala Gln Arg Thr
 1515 1520 1525

TCT CTC TCC AAG TCC TAC ATC CTG TAT GAC CTG CAG GAA GCC ACC TGG 5081
 Ser Leu Ser Lys Ser Tyr Ile Leu Tyr Asp Leu Gln Glu Ala Thr Trp
 1530 1535 1540

TAT GAG CTG CAG ATG CGG GTG TGC AAC AGT GCG GGC TGC GCG GAG AAG 5129
 Tyr Glu Leu Gln Met Arg Val Cys Asn Ser Ala Gly Cys Ala Glu Lys
 1545 1550 1555

CAG GCT AAA GAG GCT GCG AGA TGC AAA GAG TTT AGC TGAAATGCTC 5175
 Gln Ala Lys Glu Ala Ala Arg Cys Lys Glu Phe Ser
 1560 1565 1570

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CACATCGACA TACCCAGGGC TCAGCTTTTG ATTGAAGAGA GAGACACGAT GGAGACCATT 5295

GATGATCGCT CCACGGTTCT GTTGACGGAT GCTGACTTTG GAGAGGCAGC TAAGCAGAAG 5355

TCCCTGACGG TCACTCACAC GGTCCATTAC CAATCGGTGT CTCAGGCCAC TGGGCCCTTA 5415

GTGGATGTTT CAGACGCTCG GCCGGGAACG AATCCCACCA CCAGGAGGAA TGCCAAGGCT 5475

GGGCCCACAG CGAGAAACCG CTATGCCAGC CAGTGGACCC TCAACCGACC CCACCCCACC 5535

ATCTCAGCAC ACACCCTCAC CACAGACTGG AGGCTGCCAA CACCCAGGGC TGCAGGATCA 5595

GTAGACAAAG AGAGCGACAG TTACAGCGTC AGCCCCTCGC AAGACACAGA TCGAGCAAGA 5655

(2) INFORMATION FOR SEQ ID NO:11:

(A) LENGTH: 1571 amino acids

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Met 1	Trp	Ile	Leu	Ala 5	Leu	Ser	Leu	Phe	Gln 10	Ser	Phe	Ala	Asn	Val 15	Phe
Ser	Glu	Asp	Leu 20	His	Ser	Ser	Leu	Tyr 25	Phe	Val	Asn	Ala	Ser 30	Leu	Gln
Glu	Val	Val 35	Phe	Ala	Ser	Thr	Thr 40	Gly	Thr	Leu	Val	Pro 45	Cys	Pro	Ala
Ala	Gly 50	Ile	Pro	Pro	Val	Thr 55	Leu	Arg	Trp	Tyr	Leu 60	Ala	Thr	Gly	Glu
Glu 65	Ile	Tyr	Asp	Val	Pro 70	Gly	Ile	Arg	His	Val 75	His	Pro	Asn	Gly	Thr 80
Leu	Gln	Ile	Phe	Pro 85	Phe	Pro	Pro	Ser	Ser 90	Phe	Ser	Thr	Leu	Ile 95	His
Asp	Asn	Thr	Tyr 100	Tyr	Cys	Thr	Ala	Glu 105	Asn	Pro	Ser	Gly	Lys 110	Ile	Arg
Ser	Gln	Asp 115	Val	His	Ile	Lys	Ala 120	Val	Leu	Arg	Glu	Pro 125	Tyr	Thr	Val

Arg Val Glu Asp Gln Lys Thr Met Arg Gly Asn Val Ala Val Phe Lys
 130 135 140
 Cys Ile Ile Pro Ser Ser Val Glu Ala Tyr Ile Thr Val Val Ser Trp
 145 150 155 160
 Glu Lys Asp Thr Val Ser Leu Val Ser Gly Ser Arg Phe Leu Ile Thr
 165 170 175
 Ser Thr Gly Ala Leu Tyr Ile Lys Asp Val Gln Asn Glu Asp Gly Leu
 180 185 190
 Tyr Asn Tyr Arg Cys Ile Thr Arg His Arg Tyr Thr Gly Glu Thr Arg
 195 200 205
 Gln Ser Asn Ser Ala Arg Leu Phe Val Ser Asp Pro Ala Asn Ser Ala
 210 215 220
 Pro Ser Ile Leu Asp Gly Phe Asp His Arg Lys Ala Met Ala Gly Gln
 225 230 235 240
 Arg Val Glu Leu Pro Cys Lys Ala Leu Gly His Pro Glu Pro Asp Tyr
 245 250 255
 Arg Trp Leu Lys Asp Asn Met Pro Leu Glu Leu Ser Gly Arg Phe Gln
 260 265 270
 Lys Thr Val Thr Gly Leu Leu Ile Glu Asn Ile Arg Pro Ser Asp Ser
 275 280 285
 Gly Ser Tyr Val Cys Glu Val Ser Asn Arg Tyr Gly Thr Ala Lys Val
 290 295 300
 Ile Gly Arg Leu Tyr Val Lys Gln Pro Leu Lys Ala Thr Ile Ser Pro
 305 310 315 320
 Arg Lys Val Lys Ser Ser Val Gly Ser Gln Val Ser Leu Ser Cys Ser
 325 330 335
 Val Thr Gly Thr Glu Asp Gln Glu Leu Ser Trp Tyr Arg Asn Gly Glu
 340 345 350
 Ile Leu Asn Pro Gly Lys Asn Val Arg Ile Thr Gly Ile Asn His Glu
 355 360 365
 Asn Leu Ile Met Asp His Met Val Lys Ser Asp Gly Gly Ala Tyr Gln
 370 375 380
 Cys Phe Val Arg Lys Asp Lys Leu Ser Ala Gln Asp Tyr Val Gln Val
 385 390 395 400
 Val Leu Glu Asp Gly Thr Pro Lys Ile Ile Ser Ala Phe Ser Glu Lys
 405 410 415
 Val Val Ser Pro Ala Glu Pro Val Ser Leu Met Cys Asn Val Lys Gly
 420 425 430
 Thr Pro Leu Pro Thr Ile Thr Trp Thr Leu Asp Asp Asp Pro Ile Leu
 435 440 445
 Lys Gly Gly Ser His Arg Ile Ser Gln Met Ile Thr Ser Glu Gly Asn
 450 455 460

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Leu Gly Glu Ile Lys Asn Ile Thr Thr Thr Gln Pro Ser Leu Glu Leu
 1140 1145 1150
 Asp Gly Leu Glu Lys Tyr Thr Asn Tyr Ser Ile Gln Val Leu Ala Phe
 1155 1160 1165
 Thr Arg Ala Gly Asp Gly Val Arg Ser Glu Gln Ile Phe Thr Arg Thr
 1170 1175 1180
 Lys Glu Asp Val Pro Gly Pro Pro Ala Gly Val Lys Ala Ala Ala Ala
 1185 1190 1195 1200
 Ser Ala Ser Met Val Phe Val Ser Trp Leu Pro Pro Leu Lys Leu Asn
 1205 1210 1215
 Gly Ile Ile Arg Lys Tyr Thr Val Phe Cys Ser His Pro Tyr Pro Thr
 1220 1225 1230
 Val Ile Ser Glu Phe Glu Ala Ser Pro Asp Ser Phe Ser Tyr Arg Ile
 1235 1240 1245
 Pro Asn Leu Ser Arg Asn Arg Gln Tyr Ser Val Trp Val Val Ala Val
 1250 1255 1260
 Thr Ser Ala Gly Arg Gly Asn Ser Ser Glu Ile Ile Thr Val Glu Pro
 1265 1270 1275 1280
 Leu Ala Lys Ala Pro Ala Arg Ile Leu Thr Phe Ser Gly Thr Val Thr
 1285 1290 1295
 Thr Pro Trp Met Lys Asp Ile Val Leu Pro Cys Lys Ala Val Gly Asp
 1300 1305 1310
 Pro Ser Pro Ala Val Lys Trp Met Lys Asp Ser Asn Gly Thr Pro Ser
 1315 1320 1325
 Leu Val Thr Ile Asp Gly Arg Arg Ser Ile Phe Ser Asn Gly Ser Phe
 1330 1335 1340
 Ile Ile Arg Thr Val Lys Ala Glu Asp Ser Gly Tyr Tyr Ser Cys Ile
 1345 1350 1355 1360
 Ala Asn Asn Asn Trp Gly Ser Asp Glu Ile Ile Leu Asn Leu Gln Val
 1365 1370 1375
 Gln Val Pro Pro Asp Gln Pro Arg Leu Thr Val Ser Lys Thr Thr Ser
 1380 1385 1390
 Ser Ser Ile Thr Leu Ser Trp Leu Pro Gly Asp Asn Gly Gly Ser Ser
 1395 1400 1405
 Ile Arg Gly Tyr Ile Leu Gln Tyr Ser Glu Asp Asn Ser Glu Gln Trp
 1410 1415 1420
 Gly Ser Phe Pro Ile Ser Pro Ser Glu Arg Ser Tyr Arg Leu Glu Asn
 1425 1430 1435 1440
 Leu Lys Cys Gly Thr Trp Tyr Lys Phe Thr Leu Thr Ala Gln Asn Gly
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 Val Gly Pro Gly Arg Ile Ser Glu Ile Ile Glu Ala Lys Thr Leu Gly
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Lys Glu Pro Gln Phe Ser Lys Glu Gln Glu Leu Phe Ala Ser Ile Asn
 1475 1480 1485
 Thr Thr Arg Val Arg Leu Asn Leu Ile Gly Trp Asn Asp Gly Gly Cys
 1490 1495 1500
 Pro Ile Thr Ser Phe Thr Leu Glu Tyr Arg Pro Phe Gly Thr Thr Val
 1505 1510 1515 1520
 Trp Thr Thr Ala Gln Arg Thr Ser Leu Ser Lys Ser Tyr Ile Leu Tyr
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 Asp Leu Gln Glu Ala Thr Trp Tyr Glu Leu Gln Met Arg Val Cys Asn
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 Ser Ala Gly Cys Ala Glu Lys Gln Ala Lys Glu Ala Ala Arg Cys Lys
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 Glu Phe Ser
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That which is claimed is:

1. Isolated nucleic acid encoding a mammalian DS-CAM member of the Immunoglobulin (Ig) superfamily of proteins, or a fragment thereof, wherein said DS-CAM
5 comprises at least 7 Ig-like domains.

2. Isolated nucleic acid according to claim 1, wherein said nucleic acid, or fragments thereof, is selected from:

- 10 (a) DNA encoding the amino acid sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9,
(b) DNA that hybridizes to the DNA of (a) under moderately stringent conditions, wherein said DNA
15 encodes biologically active DS-CAM, or
(c) DNA degenerate with respect to either (a) or (b) above, wherein said DNA encodes biologically active DS-CAM.

3. A nucleic acid according to claim 2, wherein
20 said nucleic acid hybridizes under high stringency conditions to the DS-CAM coding portion of nucleotides SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10.

4. A nucleic acid according to claim 2, wherein
25 the nucleotide sequence of said nucleic acid is substantially the same as that set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10.

5. A nucleic acid according to claim 2, wherein
the nucleotide sequence of said nucleic acid is the same
30 as that set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10.

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6. A nucleic acid according to claim 2, wherein said nucleic acid is cDNA.

7. A vector containing the nucleic acid of claim 2.

5 8. Recombinant cells containing the nucleic acid of claim 2.

9. An oligonucleotide comprising at least 15 nucleotides capable of specifically hybridizing with a sequence of nucleic acids of the nucleotide sequence set
10 forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9 or SEQ ID NO:10.

10. An oligonucleotide according to claim 9, wherein said oligonucleotide is labeled with a detectable marker.

15 11. An antisense oligonucleotide capable of specifically binding to mRNA encoded by said nucleic acid according to claim 2.

12. A kit for detecting the presence of the DS-CAM cDNA sequence comprising at least one oligonucleotide
20 according to claim 10.

13. An isolated DS-CAM protein comprising at least 7 Ig-like domains.

14. A DS-CAM protein according to claim 13, further characterized by being expressed in a significantly
25 higher amount in brain versus lung, liver or kidney.

15. A DS-CAM protein according to claim 13, wherein the amino acid sequence of said protein comprises substantially the same protein sequence set forth in

SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9.

16. A DS-CAM protein according to claim 15 comprising the same amino acid sequence as the protein
5 sequence set forth in SEQ ID NO:2 or SEQ ID NO:11, or the DS-CAM coding region of SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9.

17. A DS-CAM protein according to claim 13, wherein said protein is encoded by a nucleotide sequence
10 comprising substantially the same nucleotide sequence set forth in SEQ ID NO:1, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, or SEQ ID NO:10.

18. A DS-CAM protein according to claim 17, wherein said protein is encoded by a nucleotide sequence
15 comprising SEQ ID NO:1 or SEQ ID NO:10.

19. A DS-CAM protein according to claim 13, wherein said protein is encoded by a nucleotide sequence that comprises substantially the same nucleotide sequence as nucleotides 453-6185 set forth in SEQ ID NO:1,
20 nucleotides 453-5168 set forth in SEQ ID NO:10, SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9.

20. Method for expression of a DS-CAM-related protein, said method comprising culturing cells of claim 8 under conditions suitable for expression of said DS-CAM
25 protein.

21. An isolated anti-DS-CAM antibody having specific reactivity with a DS-CAM protein according to claim 13.

22. Antibody according to claim 21, wherein said
30 antibody is a monoclonal antibody.

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nucleic acid sequence derived from the nucleic acid
sequence set forth as SEQ ID NO: 1, SEQ ID NO:10,
SEQ ID NO:7, SEQ ID NO:8 or SEQ ID NO:9.

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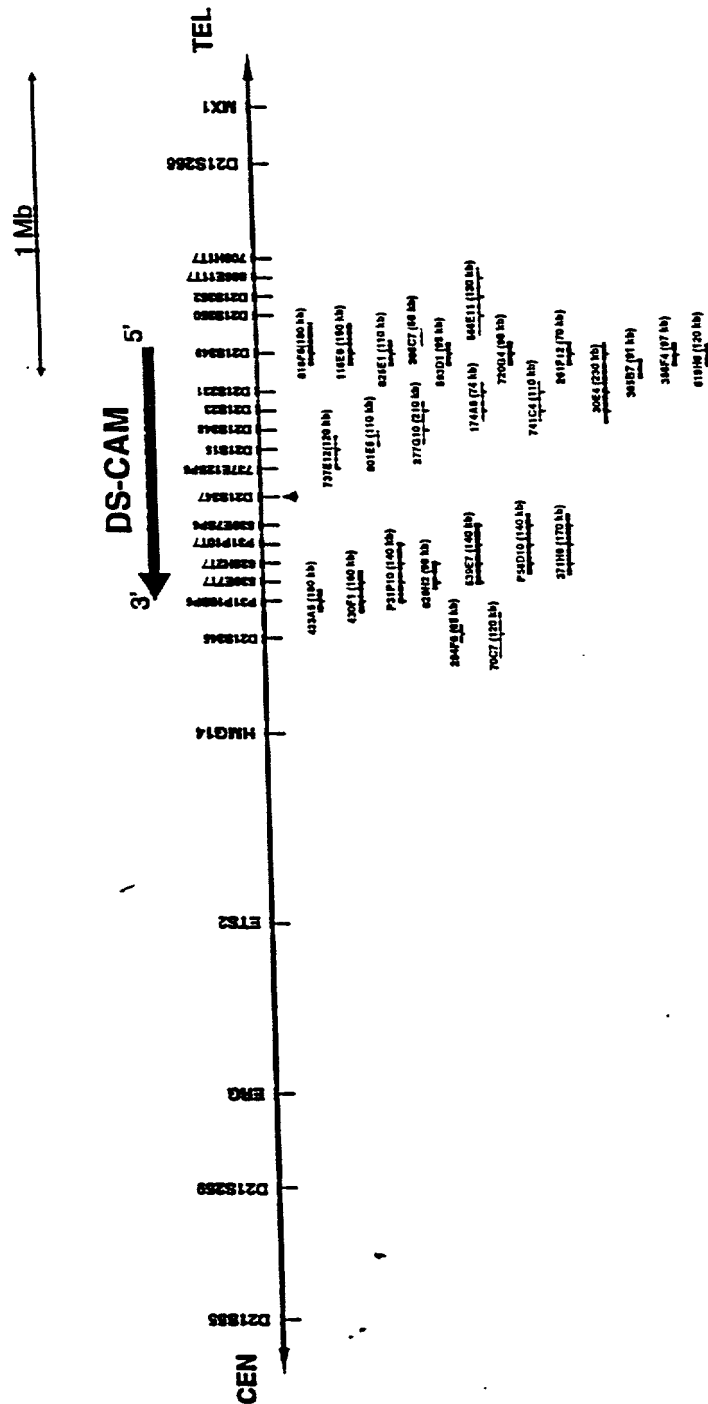


Figure 2

SEQ ID NO:2

1 MWLALSLFGSFANVFEEDLHSS

24 LYFVNASLQEVVFASTTGTLPVCPAAGIPFVTLRWYLATGEEIYDVPGRHVHPNGTLQIFFPSSSF
STLIHNDNTYYCTAEMPAGKIRSQDVHIKAVLREPY127 TVRVEDQKTRGNVAVFKCIIPSSVEAYITVSWEKDVTSLVSGSRFLITSTGALYIKDVQNEGLYN
YRCITRHRYTGETRQSNARLFVSDPANSAP228 SILDGFDRHKAMAGORVELPCALGHPEPDYRWLKDNPLELSGRFQKTVTGLLIENIRPSDSGSYVC
EVSNNRYGTAKVIGRLYVKQPLKA317 TISPRVKSSVGSQVSLSCSVTGTEDQELSWYRNGEILNPGKNVRITGINHENLIMDMVKSDDGAYQ
CFVRKDKLSAQDYVQVLEDGTPKI410 ISAFSEKVVSPAEPVSLMNVKGTPLPTITWTLDLDDPILKGGSHRISQMITSEGNVVSYLNISSQVR
DGGVYRCIANNBAGVLYQARINVRGPAS507 IAPMKNITAIAGRDTYIHCRVIGYPYSIKWYKNSNLLPFNRQVAFENNGTLKLSDVQKEVDEGETT
CNVLVQPOLSTSQSVHVTVKVPPFIQFFE604 FPFISIGQRVFTPCVVVSGDLPTITWQDGRPIPGSLGVTIDNIDFTSSLRISNLSLMHNGNTTCLA
RNEAAAVENHQSGLIVRVPKVVQPR698 DQDGIYKAVILNCIAEGYPVPTIVWKFSGAGVPPQFPIALNGRIQVLSNGSLLIKHVVEEDSGYTL
CKVSNVDGADVSKSHYLTVKIPAMITS793 YPNTTLATQGGKMSCTANGKPIIVRWEKEDRIINPEMARYLVSTKEVGEEVISTLQILPTVREDS
GFFSCNAINSYGEDRGIIQLTVQEPFD888 PPEIEIKDVKARTITLRWTMGFDGNSPITGYDIECKKSDSWDSAQRTKDVSPQLNSATIIDIHPSST
YSIRHYAKNRIGKSEPSNELTITADEAA984 PDGPPQEVHLEPISSQIRVTKAPKKHLQNGIIRGYOIGYREYSTGNGFQNIISVDTSGDSEVYTL
DNLNKFTQYGLVQACNRAGTGPPSSQEIITTTLED1087 VPSYPPEVQALATSPESISISWSTLSKEALNGILQGFVVIYWANLMDGELGEIKNTTTQPSLELDG
LEKYTNYSIQVLAFTAGDGRSEQIFTRTK1186 EDVPGPPAGVGAASASMVFSWLPPLKNGIIRKYTVFCSHPYPTVISEFEASPDSTFSYRIPLSR
NRQYSVWVAVTSAGRGNEEITVEPL1282 AKAPARILTFSGTVTFPMKDIVLPCKAVGDPSPAVKWMKDSNGTSPSLVTIDGRSIFSNQSPFIIRTV
KAEDSGYYSQIANNWGSDEIILNLQ1376 VQVPPDOPRLTVSKITSSITLSWLPDNGGSSIRGYILQYSEDNSEQWGSFPISPSERSYLENLKC
GTWYKFTLTAQNGVGPGRISEIIEAKTL1472 GKEPQFSKEQLFASINTVRNLIGWNGGCPITSFTLEYRPPGTTVMTTAQRSTLSKSYILYDLO
EATVYELQHRVCSAGCAEQANFATLNYDGSTIPLIKSVVONEEGLTNEGLK

1585 MLVTISCILVGVLLLVLLLV

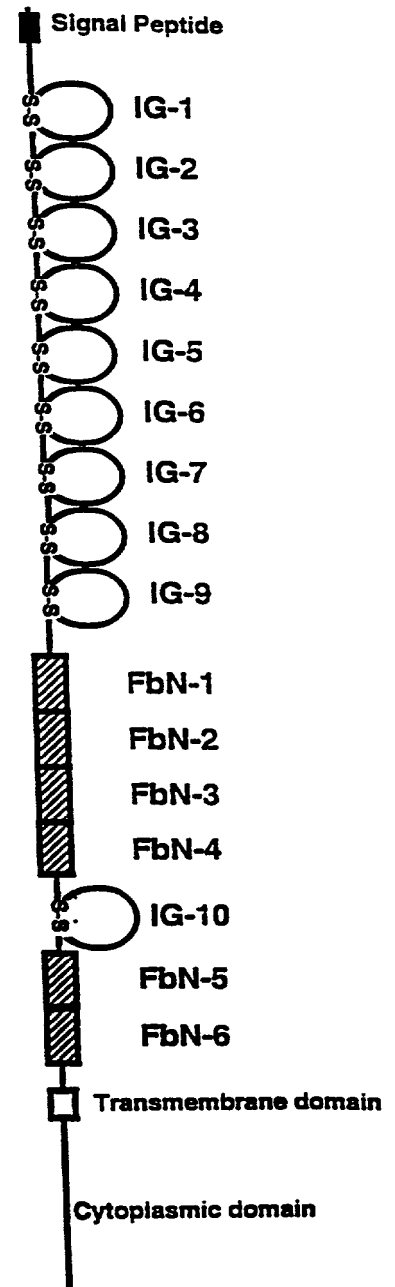
1617 RRRRREQLRLRLADAKSLAEHLMSKNTRTSDTLSKQOQTLRMHIDI PRAQLLIEERDTMETIDDRSTV
LLTDADFGAAKQKSLTVTRVYQSVSQATGPLVDVSDARPGTNPTTRNAKAGPTARNRYASQWTL
NRPHPTISANTLITDWRLPFPRAAGSVDKESDSYSVSPQDTRARSSNVSTESASSTYEELARAYER
AQMEQLRHAFTITECTISDTSSEQLTAGTNEYTDSLTSSPSESIGICRTASPPKPDGGRVMAHA
VPAIGQVTSYICLHLENTFC

Figure 3

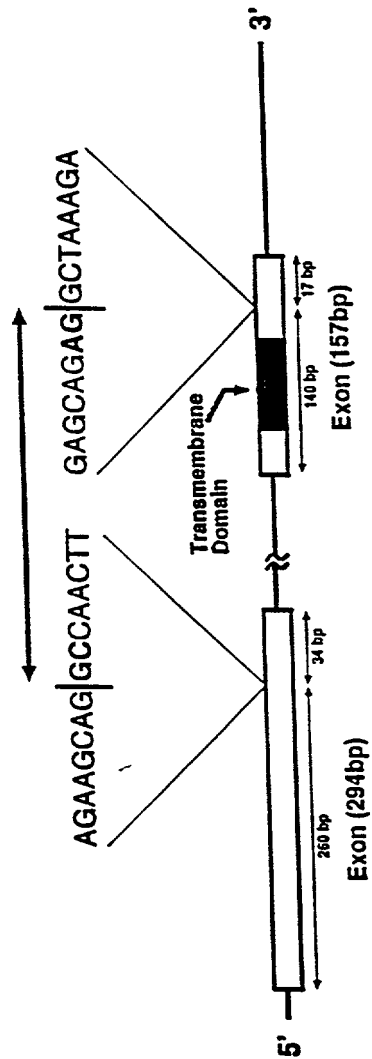


Figure 4

